



Biological Adaptation & Ageing

31 mai & 1 juin 2016

Modulation du stress oxydant

Nouveaux concepts pour de nouvelles applications

BIOCITECH, CITÉ DES ENTREPRISES DE SANTÉ ET DE BIOTECHNOLOGIES, ROMAINVILLE



Detection, quantification and identification of oxidized proteins in biological systems

Bertrand Friguet, PhD

UMR 8256 UPMC CNRS, ERL INSERM U1164

Biological Adaptation and Ageing – IBPS

University Pierre and Marie Curie
Sorbonne Universities / Paris, France

Protein modifications by oxidation and related pathways



Histidine
Cysteine
Lysine



Oxidation

Affecting all amino acids

Lysine
Arginine

Carbonyls R-C-H, R-C-R',



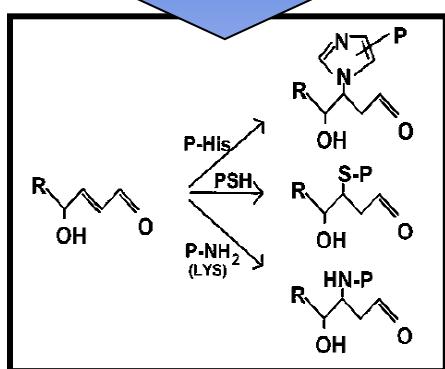
Disulfide cys-S-S-cys

Cysteine sulfenic (SO), sulfenic (SO²⁻) and sulfonic (SO³⁻) acids

Methionine sulfoxide MeSO

Cleavage of the polypeptide chain

Conjugation with
lipid peroxydation products
(e.g. HNE, MDA)



Glycation

Amadori products

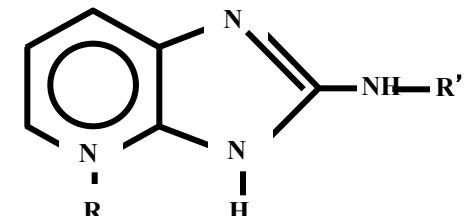
Glycoxidation

AGEs:

Carboxymethyllysine



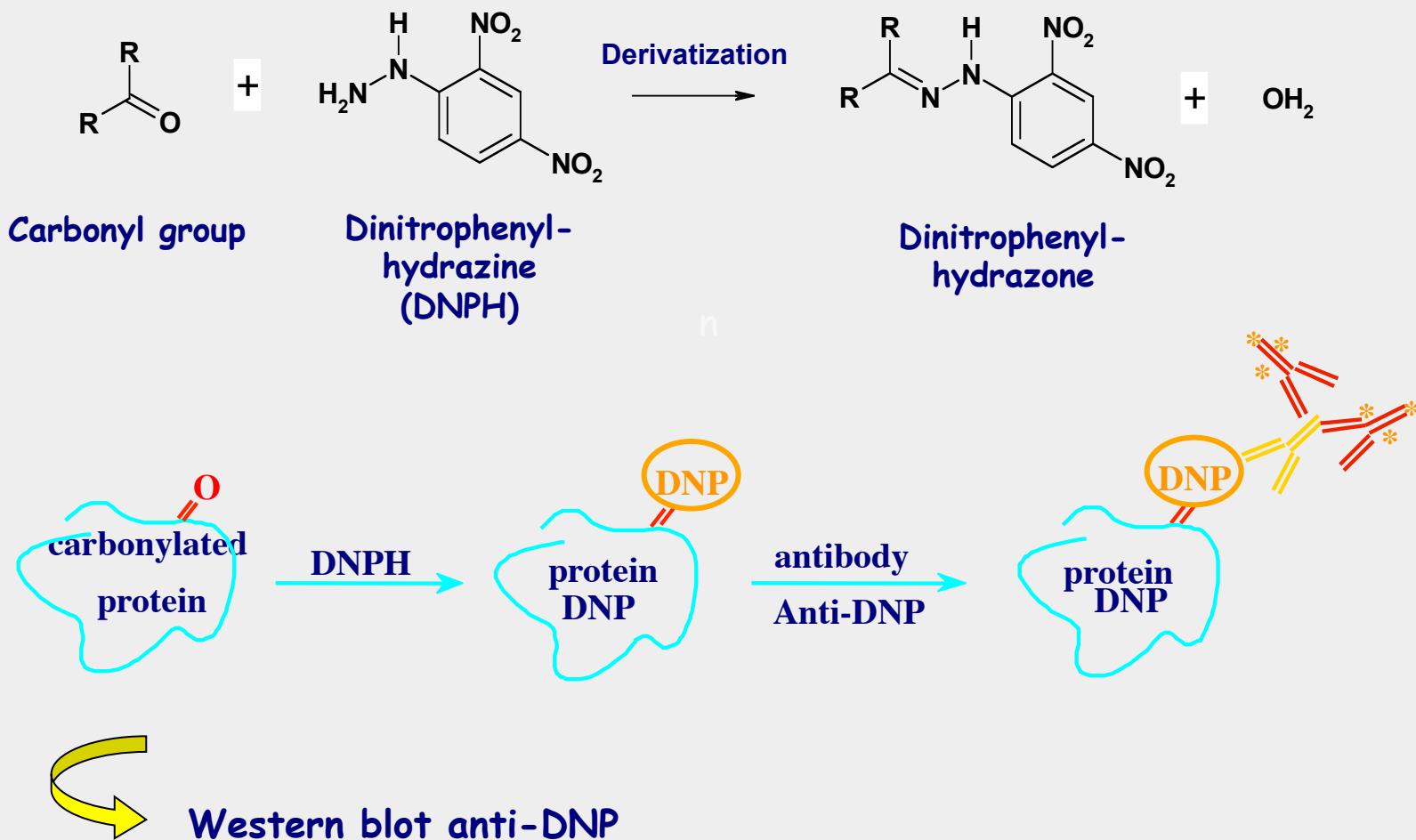
Pentosidine



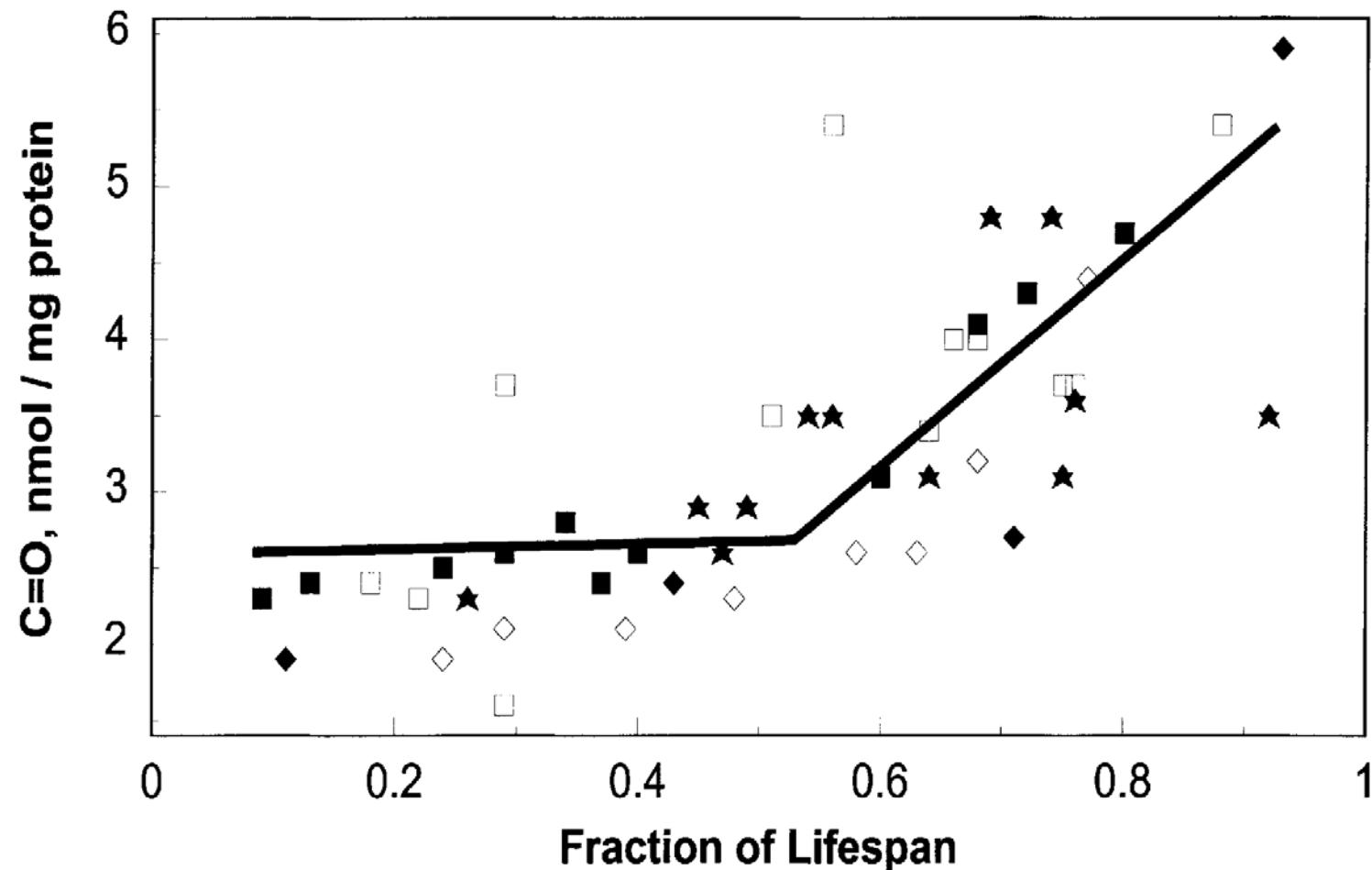
Impact of protein carbonylation

1. Biochemical activity
(loss of function)
2. Protein misfolding and aggregate formation
(gain of toxic function)
3. Cellular targeting
(protein localization)
4. Cellular metabolism and signaling
(protein-protein interaction, cascade amplification)

Immunodetection of protein carbonyl groups

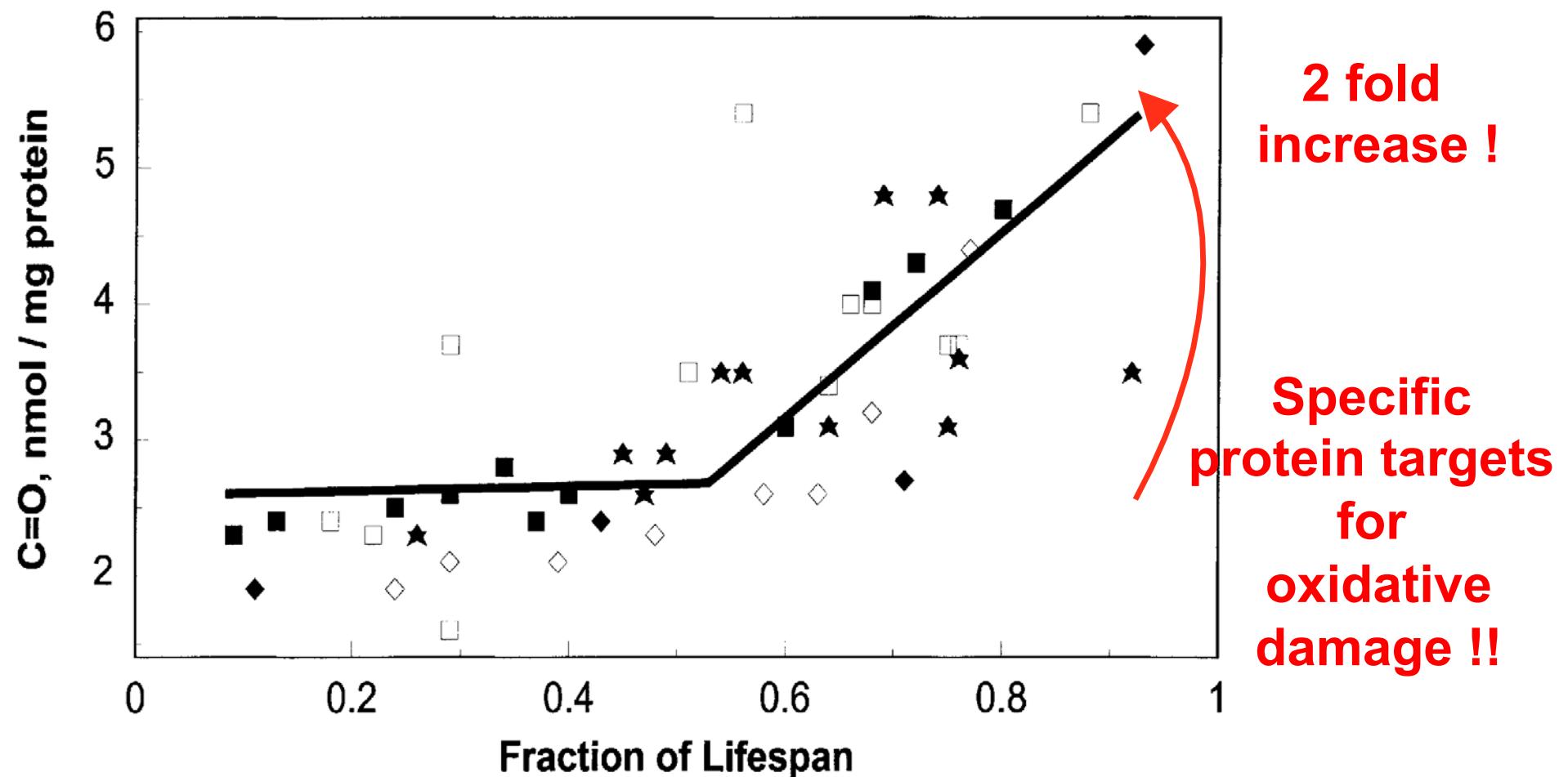


Age-related accumulation of oxidized proteins



From: Levine and Stadtman, 2001, Exp. Gerontol., 36:1495-1502
(human dermal fibroblasts, human lens, human brain, rat liver, whole fly)

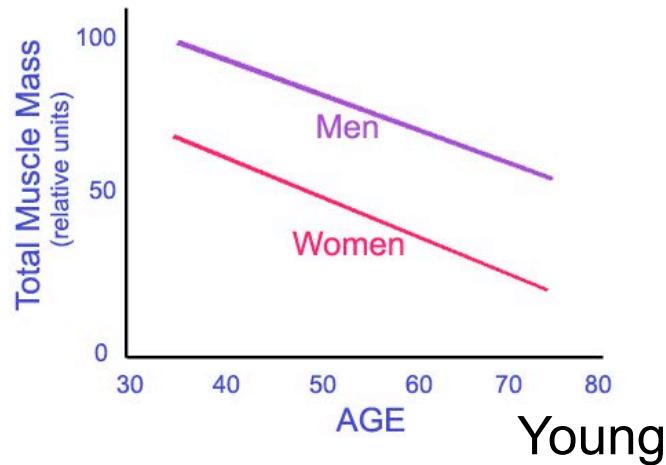
Age-related accumulation of oxidized proteins



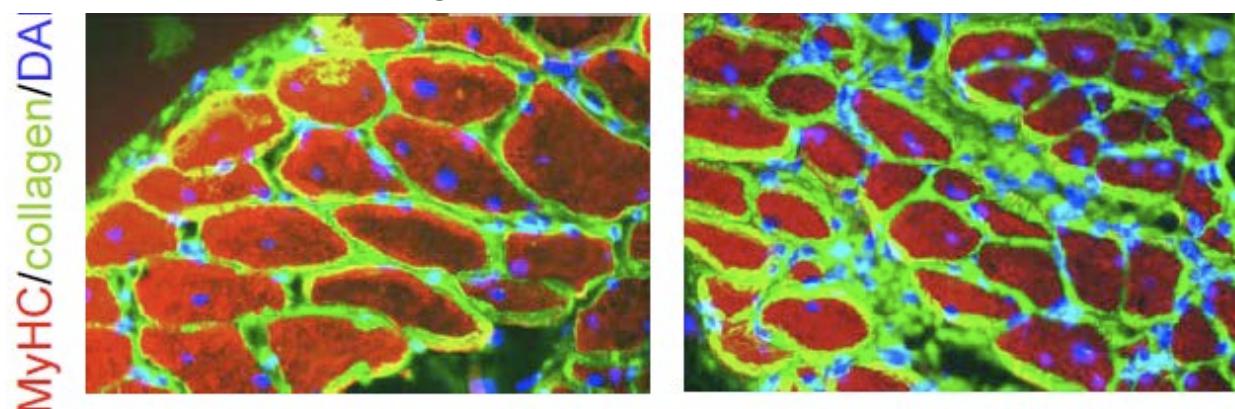
From: Levine and Stadtman, 2001, Exp. Gerontol., 36:1495-1502
(human dermal fibroblasts, human lens, human brain, rat liver, whole fly)

Skeletal Muscle Ageing

Effects of Age on Muscle Mass

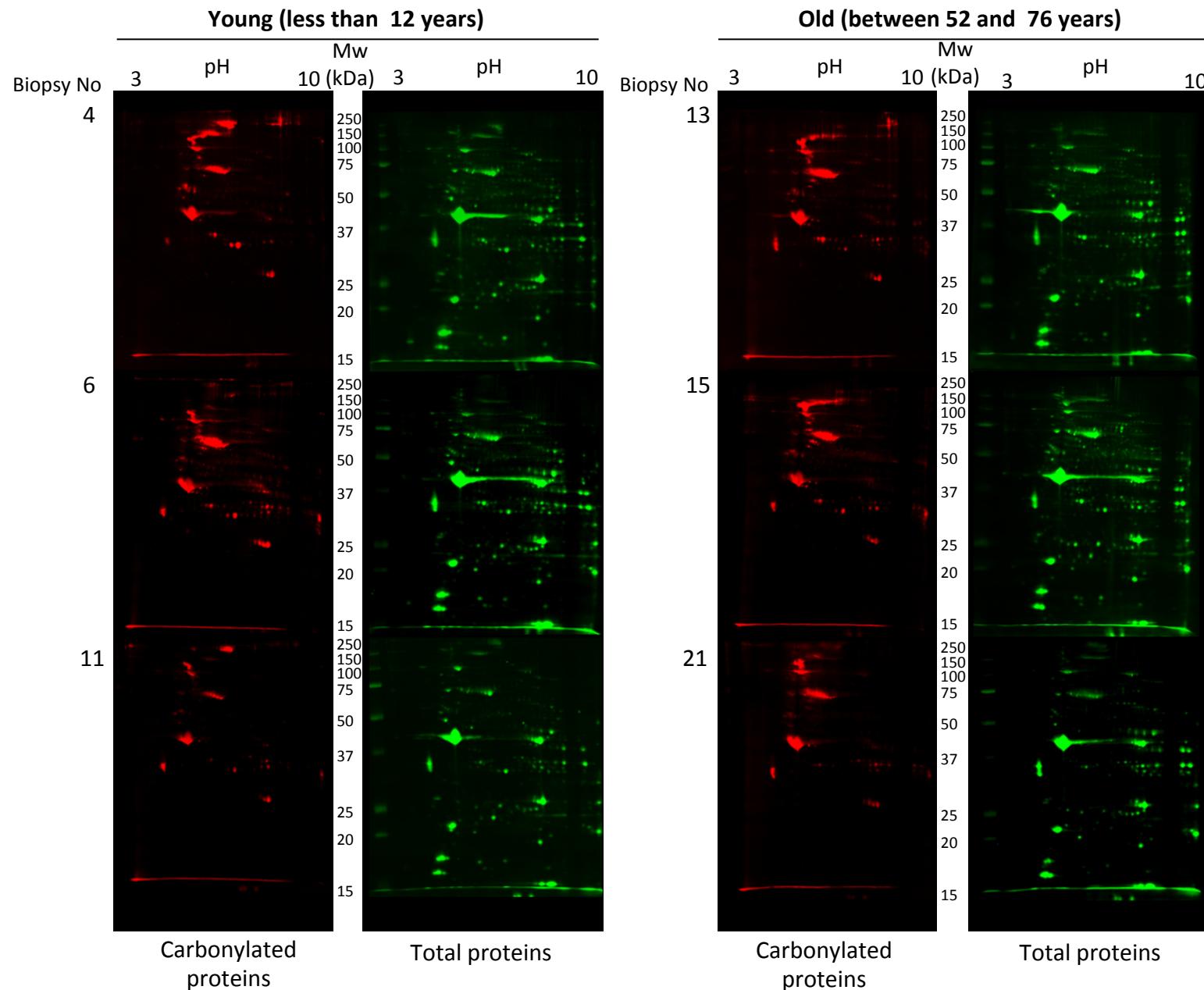


- ✓ Results in a progressive loss of mobility that decrease the quality of life
- ✓ Increasing muscle weakness is a major component of muscle ageing

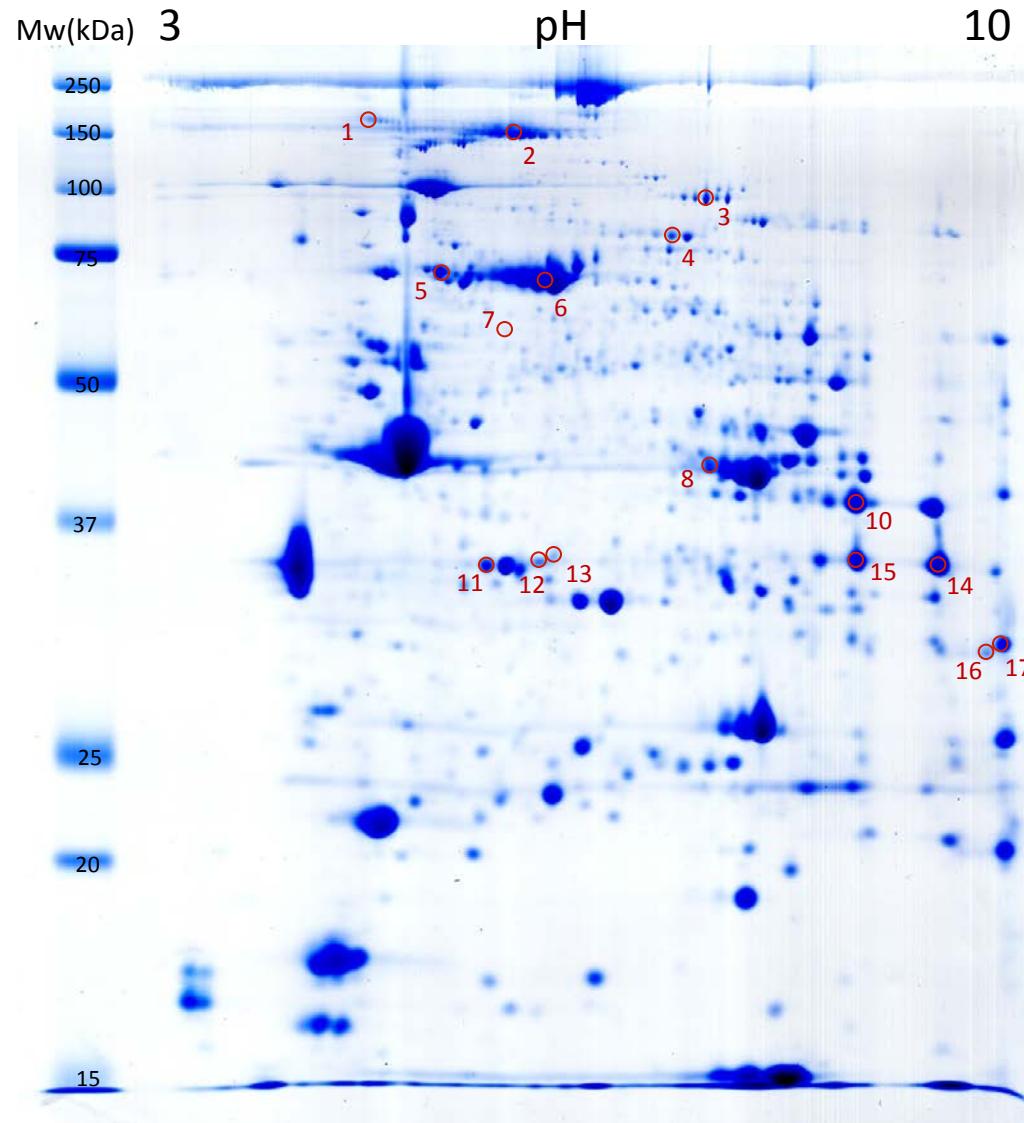


Brack AS et al. Science. (2007)

2D Oxi-proteome analysis of young and old human skeletal muscle



Coomassie blue staining of a representative 2D gel of *Rectus Abdominis* muscle: spots picked for MS identification

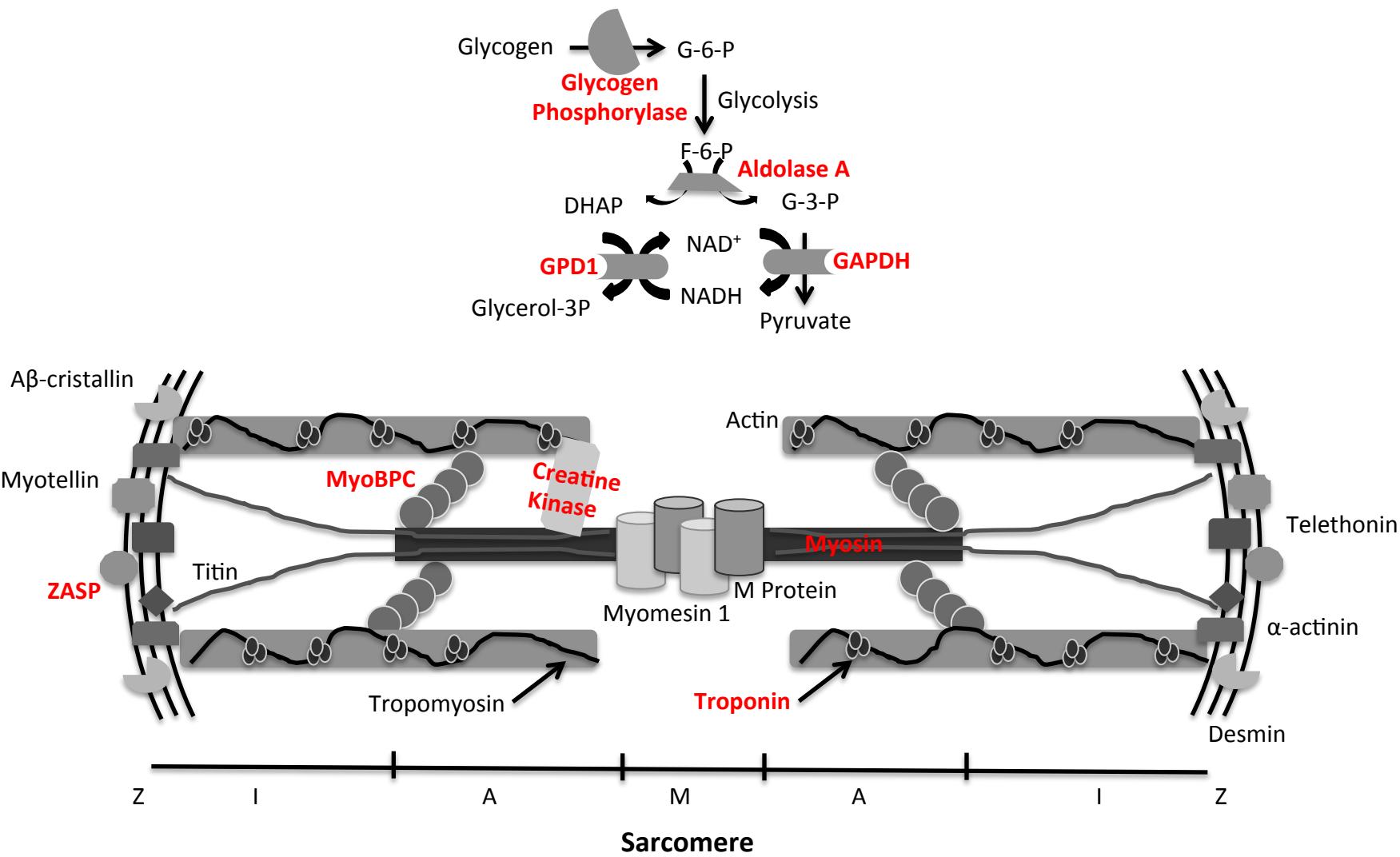


Identification of increasingly carbonylated proteins in old muscle

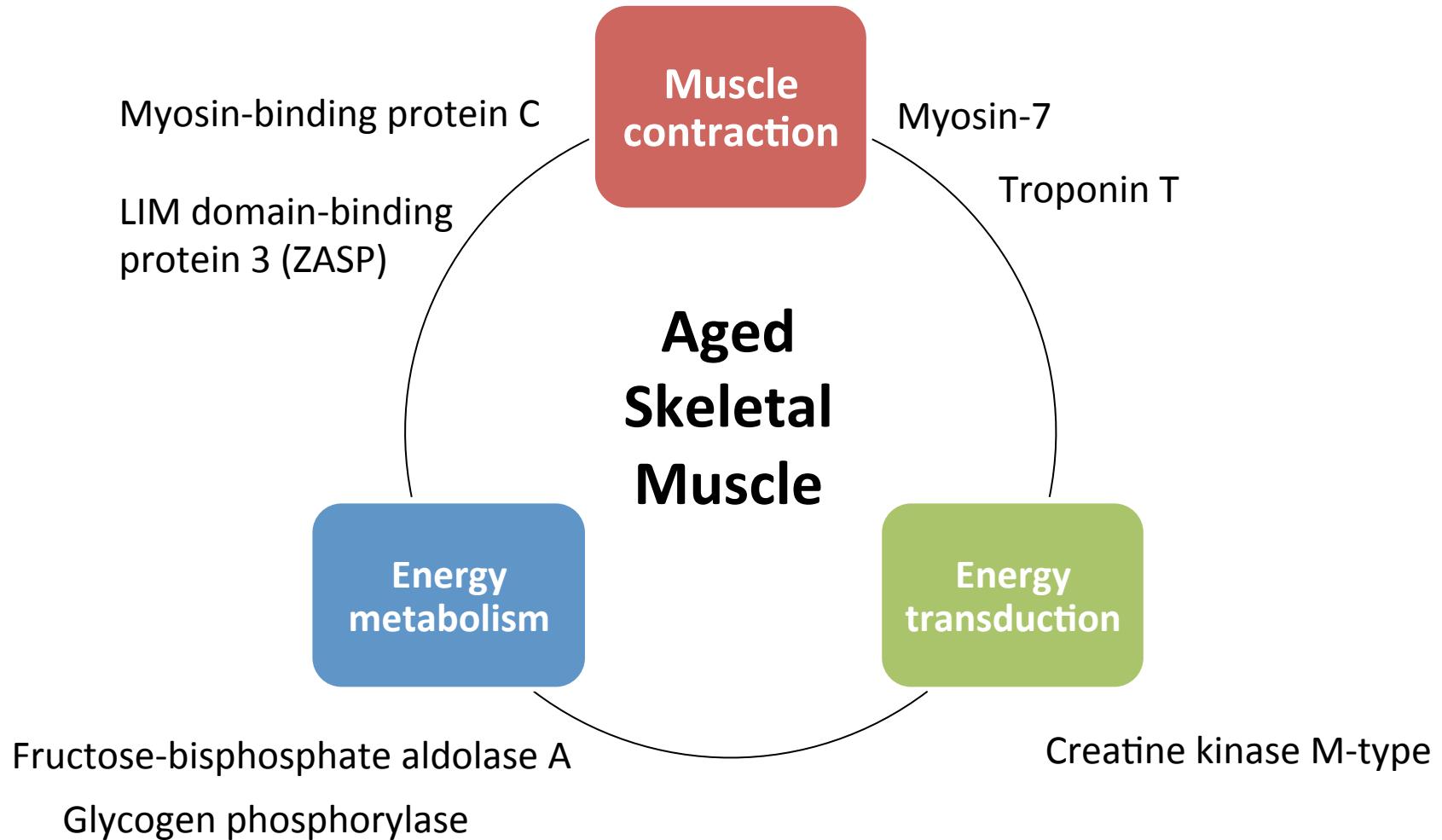
Muscle Specific Proteins

Protein name	Swiss-Prot accession no	Mascot score	Sequence coverage (%)	Theoretical mass (kDa)	Theoretical PI	RMI ratio
Collagen alpha-1(VI) chain	CO6A1_HUMAN	205	7	108	5.3	4.34
Heat shock cognate 71 kDa protein	HSP7C_HUMAN	256	15	71	5.4	1.63
Glycerol-3-phosphate dehydrogenase [NAD+], cytoplasmic (GPD1)	GPDA_HUMAN	214	22	38	5.8	1.52
Glyceraldehyde-3-phosphate dehydrogenase (GAPDH)	G3P_HUMAN	243	12	36	8.6	1.67
Glyceraldehyde-3-phosphate dehydrogenase (GAPDH)	G3P_HUMAN	424	25	36	8.6	1.68
Voltage-dependent anion-selective channel protein 1	VDAC1_HUMAN	685	39	31	8.6	1.88
Myosin-binding protein C, slow-type (MyBPC)	MYPC1_HUMAN	119	3	128	5.8	1.54
Glycogen phosphorylase, muscle form	PYGM_HUMAN	882	22	97	6.6	1.67
Myosin-7	MYH7_HUMAN	63	-	223	5.6	3.12
Creatine kinase M-type	KCRM_HUMAN	206	9	43	6.8	1.44
Creatine kinase M-type	KCRM_HUMAN	416	20	43	6.8	1.54
Fructose-bisphosphate aldolase A	ALDOA_HUMAN	611	31	39	8.3	1.65
Troponin T, slow skeletal muscle	TNNT1_HUMAN	512	22	33	5.9	1.87
Troponin T, slow skeletal muscle	TNNT1_HUMAN	244	22	33	5.9	1.42
LIM domain-binding protein 3 (ZASP)	LDB3_HUMAN	206	8	77	8.5	1.70

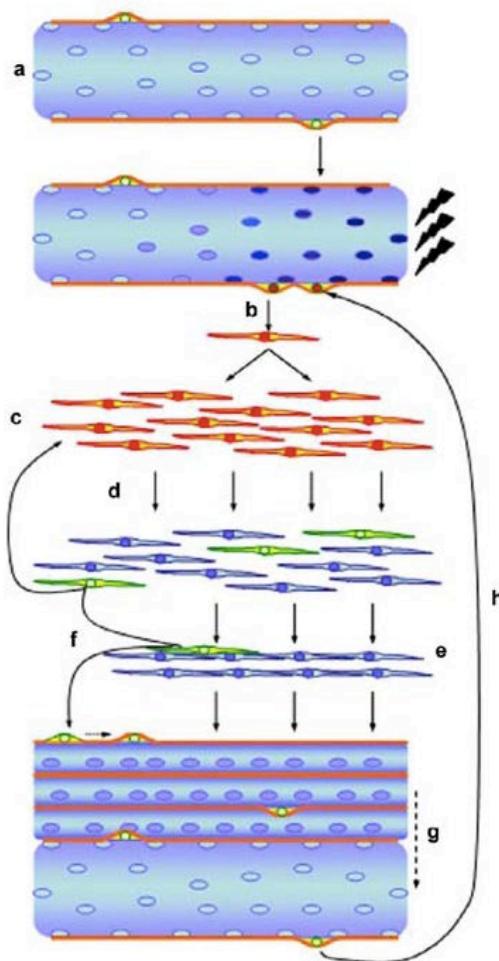
Aged *Rectus Abdominis* Muscle Oxi-Proteome



Functional grouping of skeletal muscle proteins increasingly oxidized with age

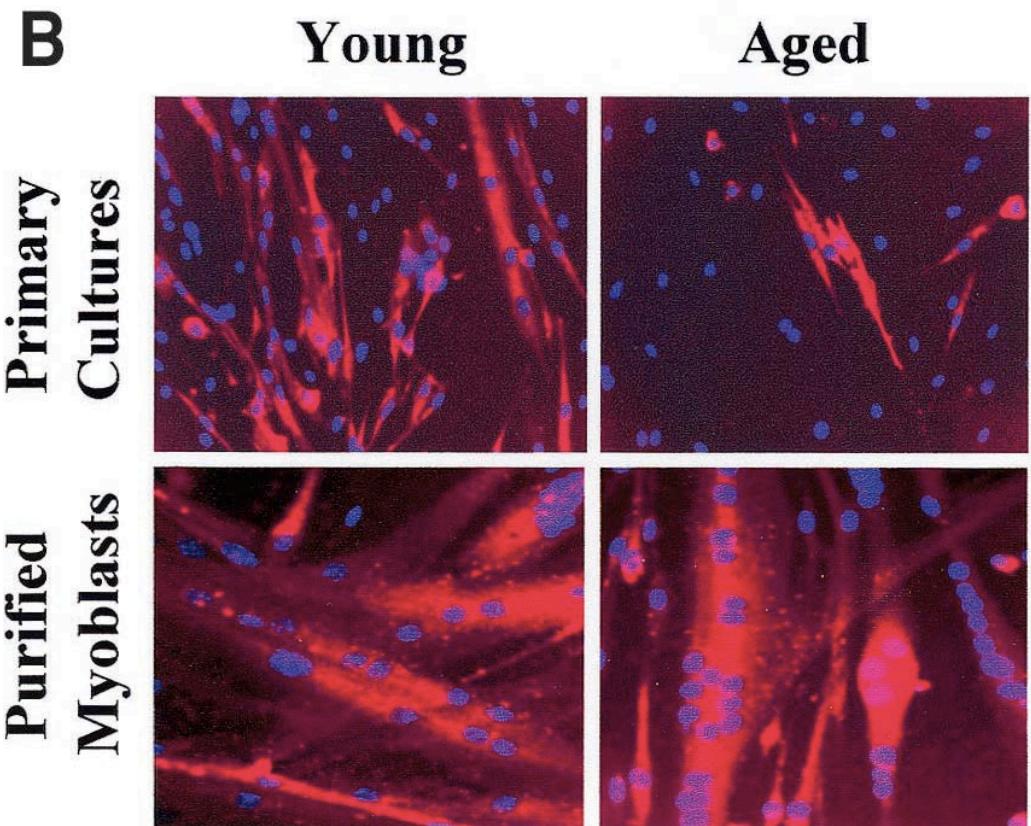


Human muscle progenitor or satellite cells are key players in skeletal muscle regeneration



Collins A et al. Cell (2005)

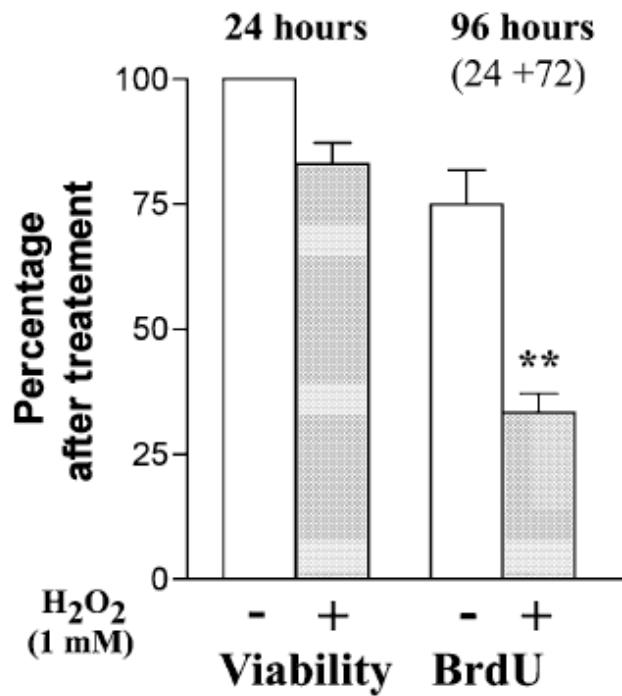
Reduced ability to proliferate and to differentiate into new myofibers during aging



Conboy I et al. Science. (2005)

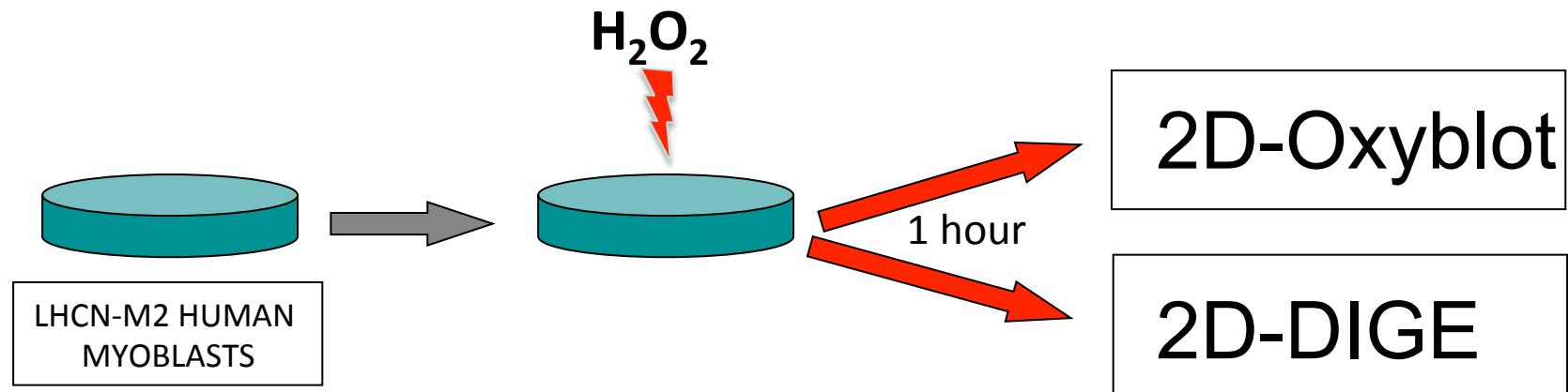
Deleterious effects of oxidative stress on human myoblasts proliferation

↓ Proliferative Capacity



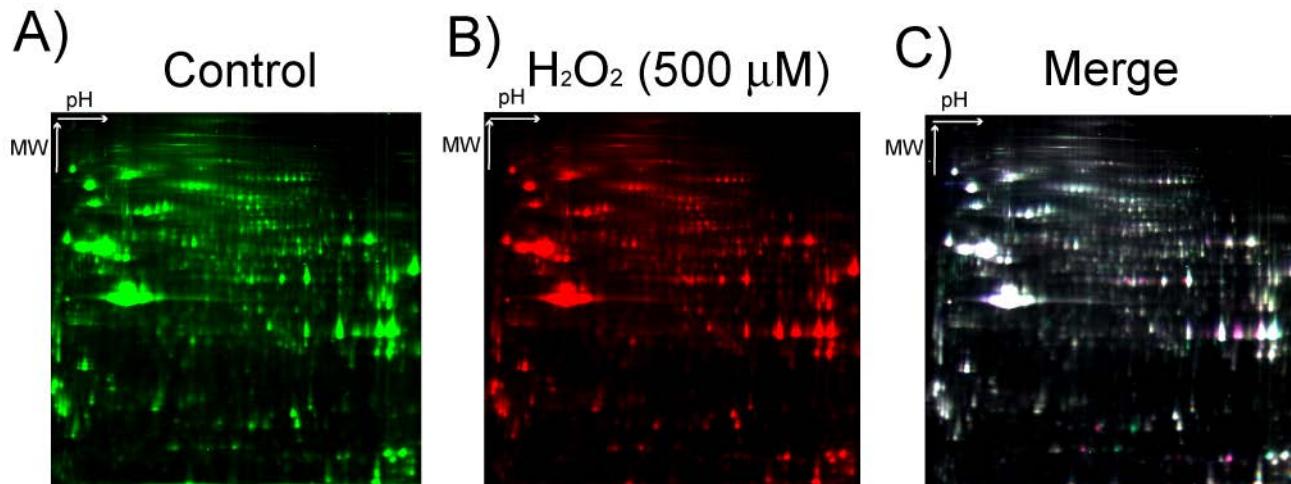
Renault et al. Exp Geront . (2002)

Oxidized protein targets in myoblasts upon acute oxidative stress

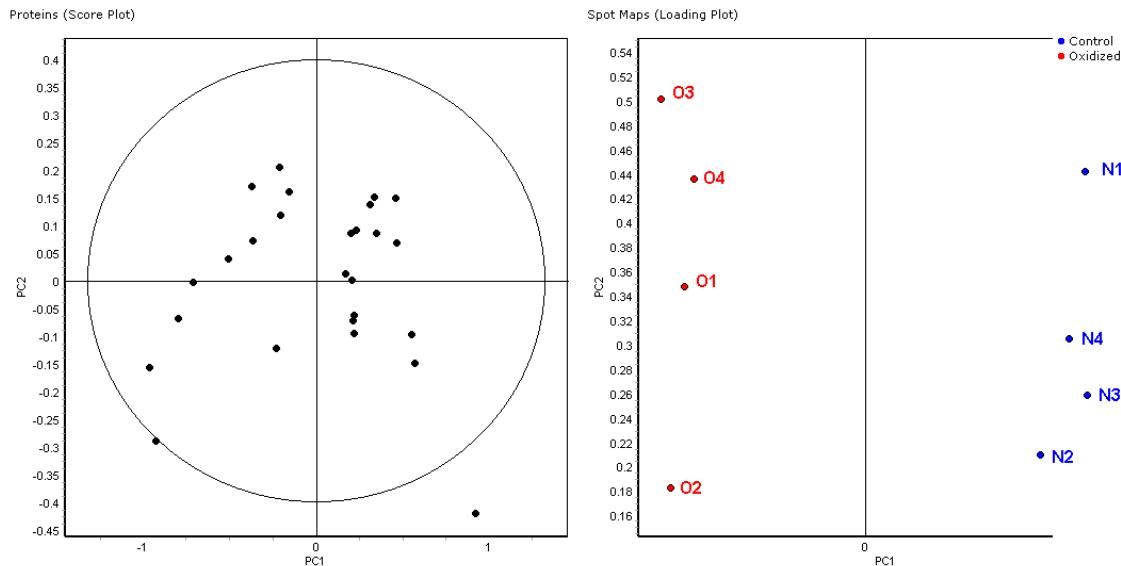


[H_2O_2] (mM)	0	0.5	1
Cell viability (%)	100	92.6±5.2	52.1±9.6*
Protein carbonyls increase (%)	-	42.6±7.6*	43.7±5.8*
Proteasome activity (% of control)	CT-L	100%±7%	66%±9%*
	T-L	100%±9%	76%±13%
	Casp-L	100%±6%	59%±5%*

Protein expression profile of human myoblasts upon oxidative stress



PCA POI - Pick List



Identification of responsive proteins upon oxidative stress

	Identified Protein	Swiss-Prot accession number	Fold change	Score Mascot	Sequence coverage (%)	Sequenced peptides	No. Of matched peptides	Theoretic Mass (Da)	Theoretic PI
Up-regulated proteins									
1	Peroxiredoxin-1	Q06830	4.64	472	46	13	4	22096	8,27
2	Peroxiredoxin-6	Q06830	4.06	345	24	10	4	22096	8,27
3	Peroxiredoxin-6	P30041	4.05	310	34	12	3	25019	6,02
4	Protein DJ-1	Q99497	3.54	128	39	-	-	19878	6,33
5	Glyceraldehyde-3-phosphate dehydrogenase	P04406	2.3	462	43	17	5	36030	8,58
7	Alpha-enolase	P06733	1.85	629	36	22	8	47139	6,99
8	Glyceraldehyde-3-phosphate dehydrogenase	P04406	1.46	793	44	27	11	36030	8,58
9	Caldesmon	Q05682	1.44	275	23	21	6	93194	5,63
10	Glyceraldehyde-3-phosphate dehydrogenase	P04406	1.41	266	31	13	4	36030	8,58
11	Adenyl cyclase associated protein 1	Q01518	1.34	118	24	12	4	51869	8,26
Down-regulated proteins									
12	Protein transport Sec23A	Q15436	-1.31	231	11	12	5	86105	8,26
13	Disulfide-isomerase A3	P30101	-1.34	798	27	23	10	56747	5,61
14	Heat shock protein β -1	P04792	-1.4	1340	36	35	18	72288	5,01
15	78 kDa glucose-regulated protein	P11021	-1.41	742	31	25	13	60864	5,43
16	Prolyl 4-hydroxylase	O15460	-1.42	1060	65	43	13	47139	6,99
17	Alpha-enolase	P06733	-1.44	342	28	17	6	61011	5,7
18	Prolyl 4-hydroxylase	P13674	-1.47	231	11	12	5	86105	8,26
19	ATP-citrate synthase	P53396	-1.6	123	13	16	2	120762	6,95
21	ATP-citrate synthase	P53396	-1.71	167	17	21	2	120762	6,95
22	ATP-citrate synthase	P53396	-2.03	95	11	15	2	120762	6,95
23	Glyceraldehyde-3-phosphate déhydrogénase	P04406	-2.09	790	46	23	8	36030	8,58
24	Péroxiredoxine-1	Q06830	-2.54	425	43	11	3	22096	8,27
25	Proteasome subunit β 3	P49720	-2.55	332	33	14	5	22933	6,13
26	Peroxiredoxin-6	P30041	-4.65	140	32	9	1	25019	6,02

Antioxidant response

Energy Metabolism

Protein Maturation

Protein Degradation

Identification of carbonylated proteins upon oxidative stress

Spot	Identified Protein	Swiss Prot accession number	Mascot Score	Sequence Coverage (%)	Seqenced Peptides	Matched peptides	Theoretic Mass (Da)	Theoretic pI	Ratio RMI
1	Elongation Factor 2	P13639	486	19	28	12	95207	6,42	2,5 ± 0,2
2	Elongation Factor 2	P13639	734	24	29	14	95207	6,42	1,8 ± 0,2
3	Elongation Factor 2	P13639	605	24	25	10	95207	6,42	7,2 ± 2,5
4	Elongation Factor 2	P13639	754	23	29	13	95207	6,42	h
5	Programmed cell death 6-interacting protein	Q8WUM4	513	31	29	10	95963	6,14	2,3 ± 1
6	Programmed cell death 6-interacting protein	Q8WUM4	361	29	30	6	95963	6,14	1,9 ± 0,1
7	Apha-glucosidase neutre AB	Q14697	675	33	39	15	103975	5,58	5,6 ± 1,5
8	Apha-glucosidase neutre AB	Q14697	675	33	39	15	103975	5,58	h
9	Gelsoline	P06396	979	29	26	11	82959	5,72	1,9 ± 0,6
10	Gelsoline	P06396	502	28	21	7	82959	5,72	1,7 ± 0,2
11	Gelsoline	P06396	185	17	14	5	82959	5,72	1,64 ± 0,1
12	Prolyl 4-hydrolase	O15460	723	37	28	11	58566	5,43	1,34 ± 0,01
13	Prolyl 4-hydrolase	O15460	723	37	28	11	58566	5,43	1,9 ± 0,1
14	Protein disulfide-isomerase A3	P30101	1030	42	35	12	54265	5,61	1,3 ± 0,1
15	Alpha-enolase	P06733	954	51	-	-	47037	6,99	1,6 ± 0,04
16	Alpha-enolase	P06733	1150	42	26	9	47037	6,99	1,7 ± 0,4
17	Protein Lupus La	P05455	135	15	11	9	46837	6,7	h
18	Proteasome subunit 10B 26S	P62333	526	24	17	8	44041	7,25	3,2 ± 0,3
19	Annexine A2	P07355	1410	46	30	15	38472	7,56	3,1 ± 0,8
20	Annexine A2	P07355	1510	56	32	19	38472	7,56	1,4 ± 0,12
21	Glyceraldehyde-3-phosphate dehydrogenase	P04406	307	32	9	3	35922	8,58	3,3 ± 0,8

Protein Synthesis

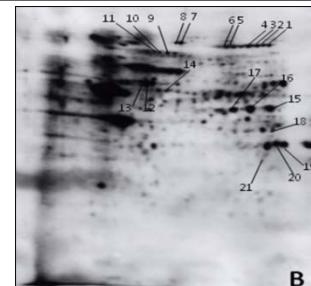
Protein Transport

Protein Maturation

Protein Degradation

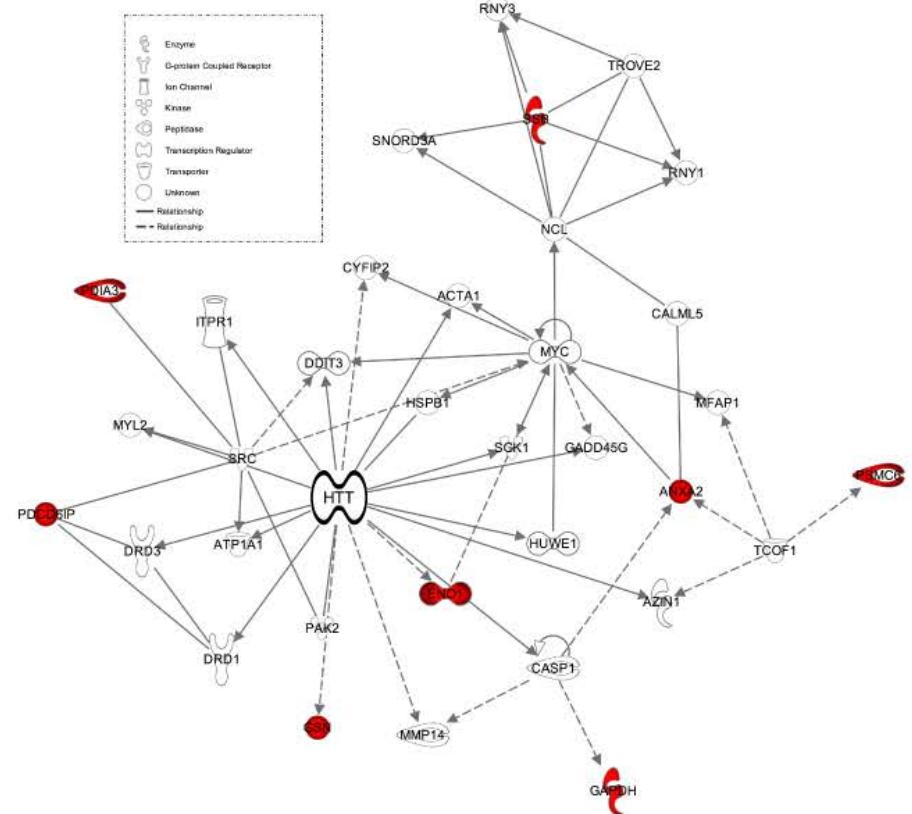
Cytoskeletal assembly

Energy Metabolism

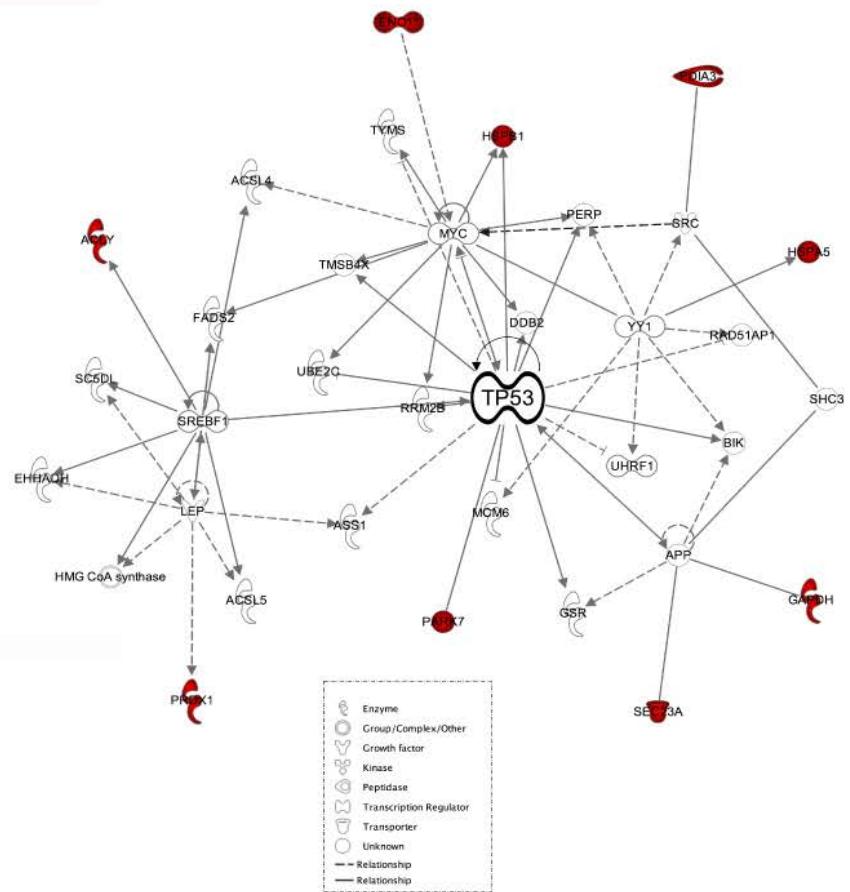


Cellular Pathway Analysis (Ingenuity™ Software)

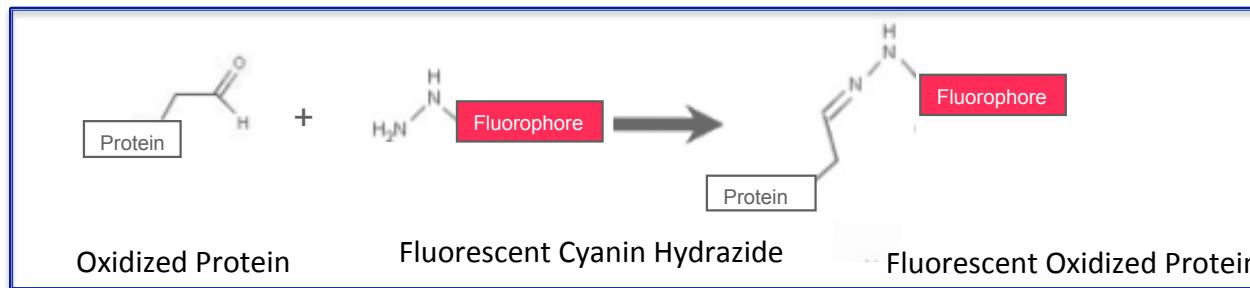
Carbonylated Proteins



Differentially Expressed Proteins



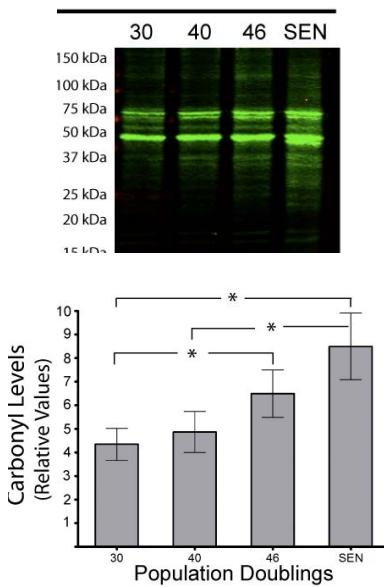
A novel approach for detecting and quantifying of carbonylated proteins: THE OXI-DIGE TECHNOLOGY



Patent: PCT/EP2012/061749 - WO2012175519

Baraibar et al., J. Proteomics, 2013

Quantification of oxidized proteins

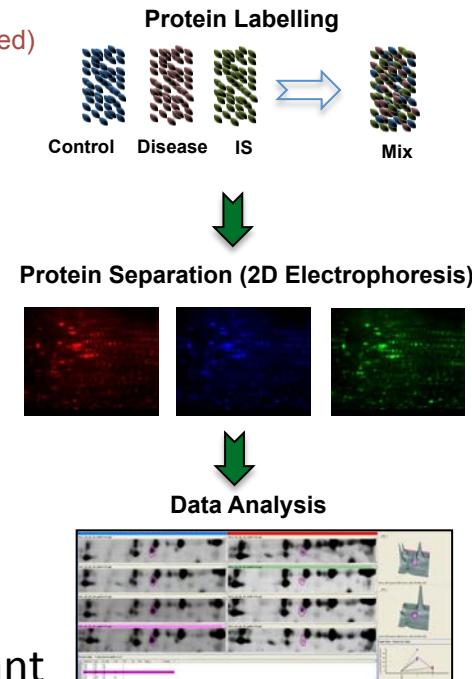


Advantages:

- ✓ Unprecedented quantification power (software aided)
- ✓ High sensitivity & accuracy
- ✓ Multiplexing: 3 comparisons per gel
- ✓ Less amount of biological material needed
- ✓ Reduced inter- and intra-assay variability
- ✓ Cost effective



Identification of specific oxidized proteins



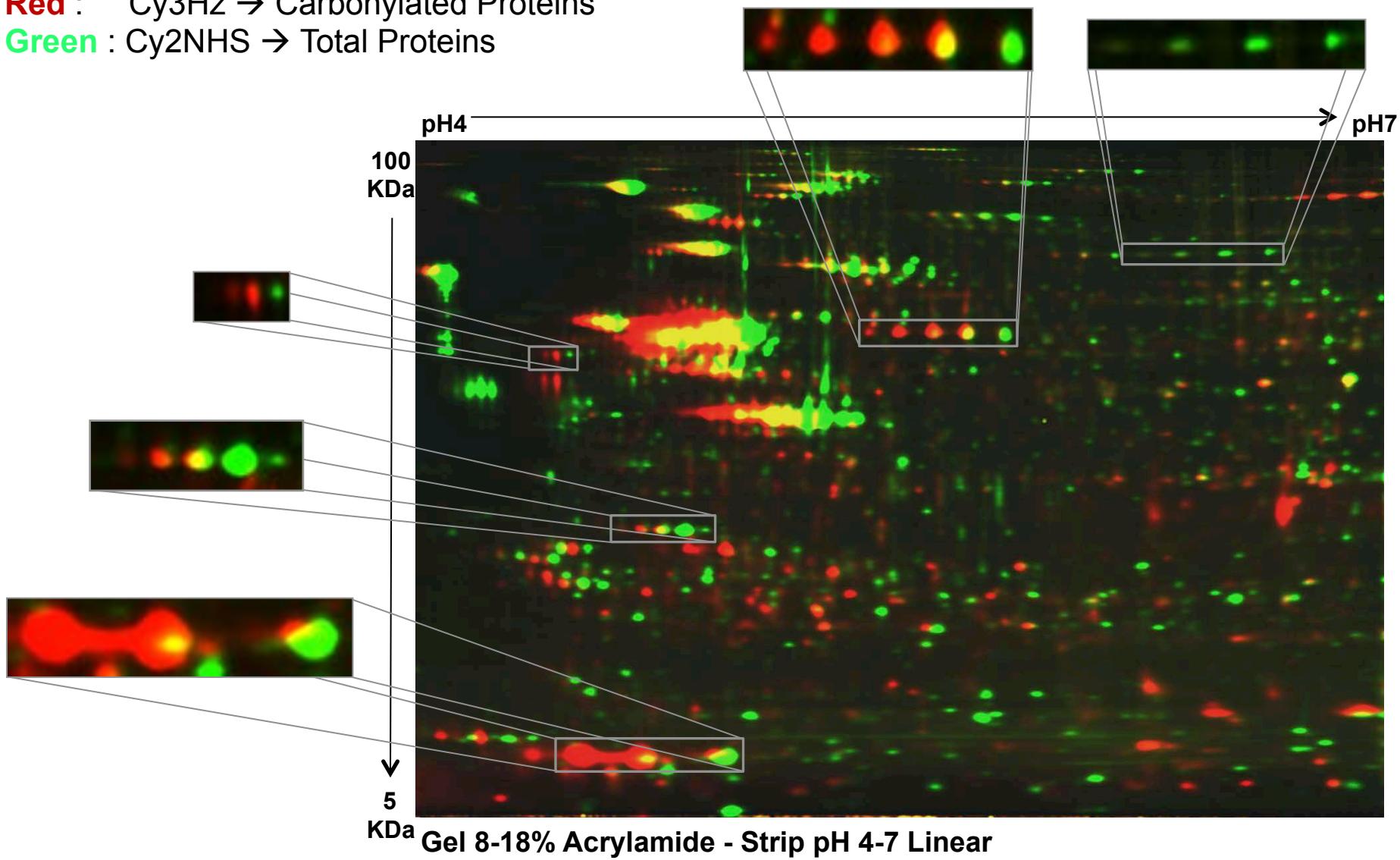
Disclosure: co-founder and scientific consultant

DETECTION OF CARBONYLATED PROTEINS AFTER 2D SEPARATION

WI-38 fibroblasts upon exposure to H_2O_2 acute oxidative stress

Red : Cy3Hz → Carbonylated Proteins

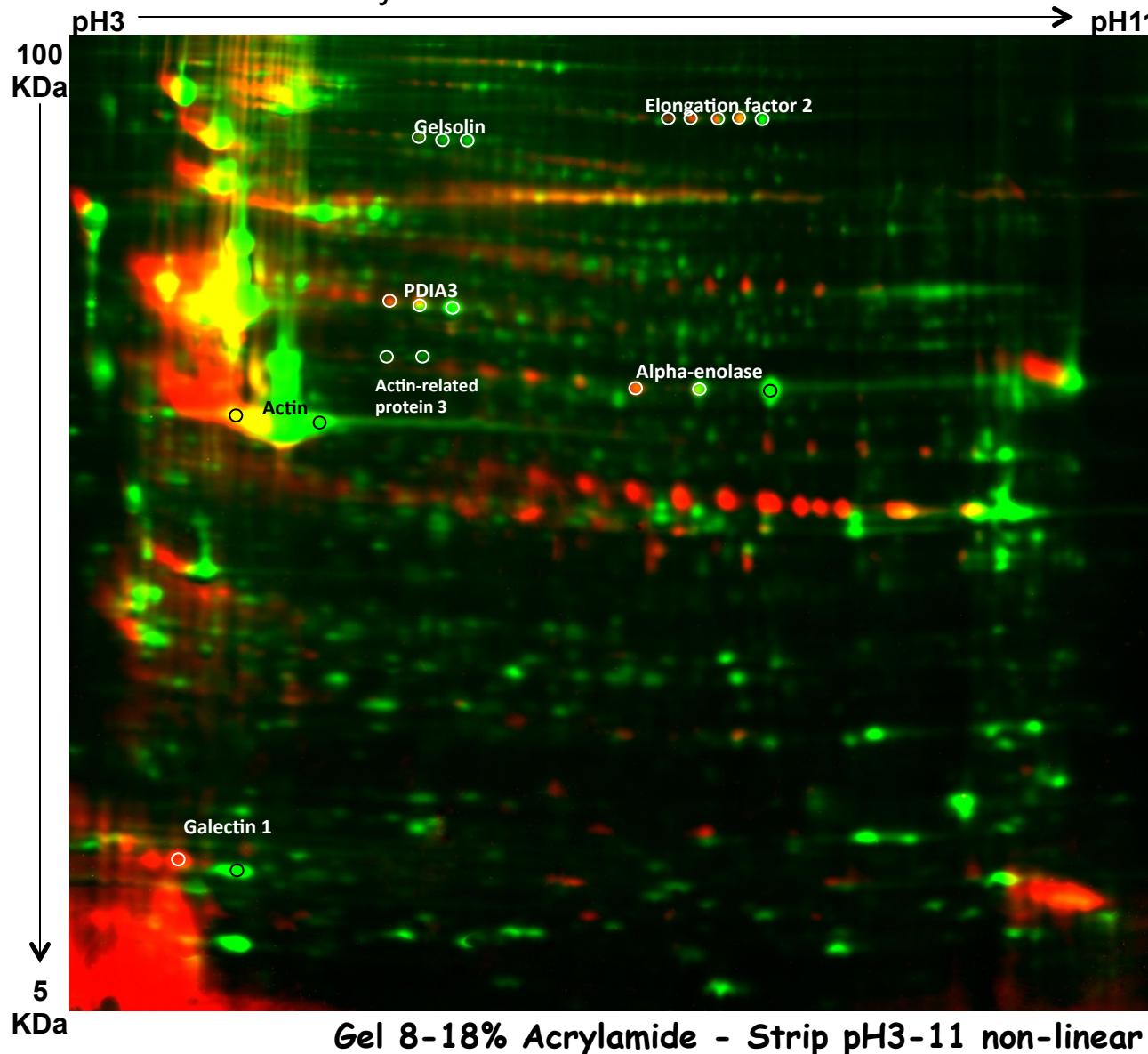
Green : Cy2NHS → Total Proteins



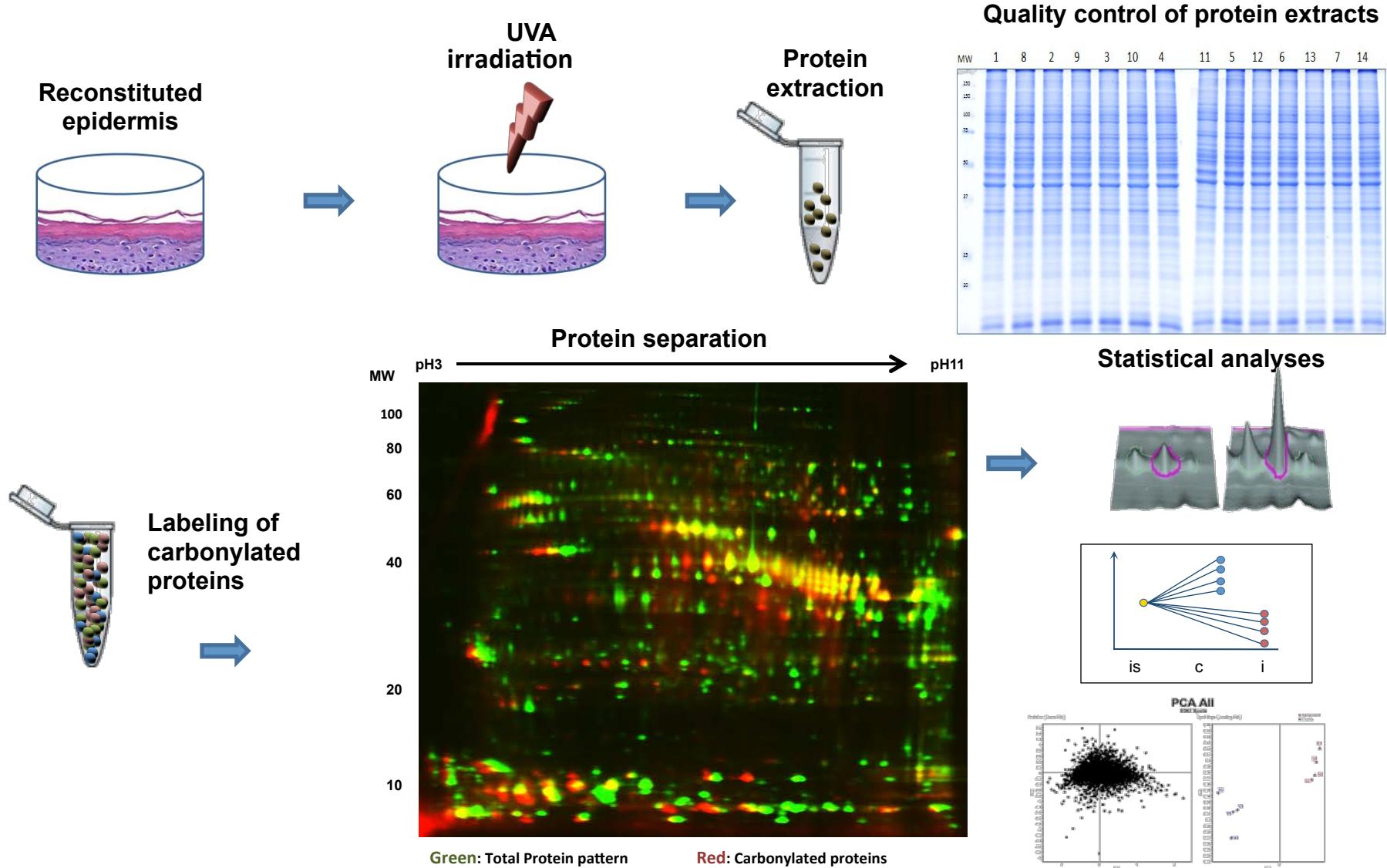
PROTEIN IDENTIFICATION BY MASS-SPECTROMETRY

Red : Cy3Hz → Carbonylated Proteins

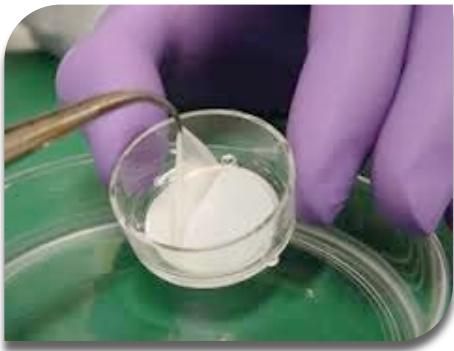
Green : Cy2NHS → Total Proteins



VALIDATION FOR PHOTO-AGING APPLICATIONS



VALIDATION FOR ACTIVE COMPOUNDS EFFICACY TESTING

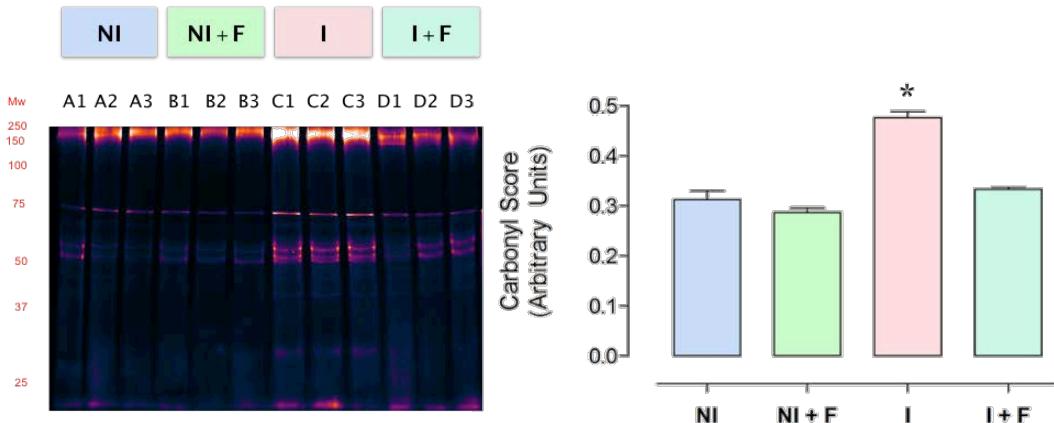


Reconstituted
skin

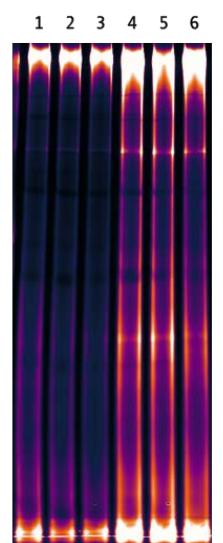


Tape Stripping
of *stratum corneum*

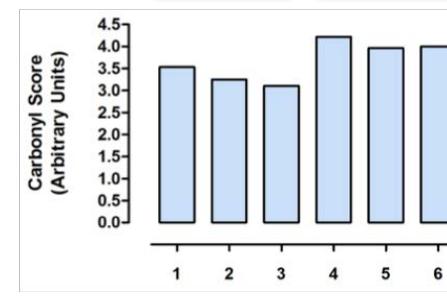
Quantification of oxidized proteins



Intrinsic Photo
Ageing Ageing

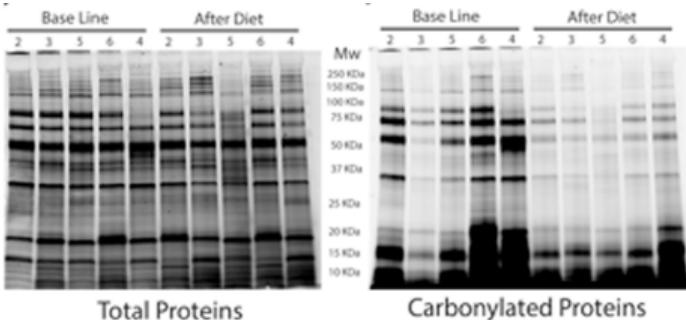


Intrinsic
Photo
Ageing Ageing



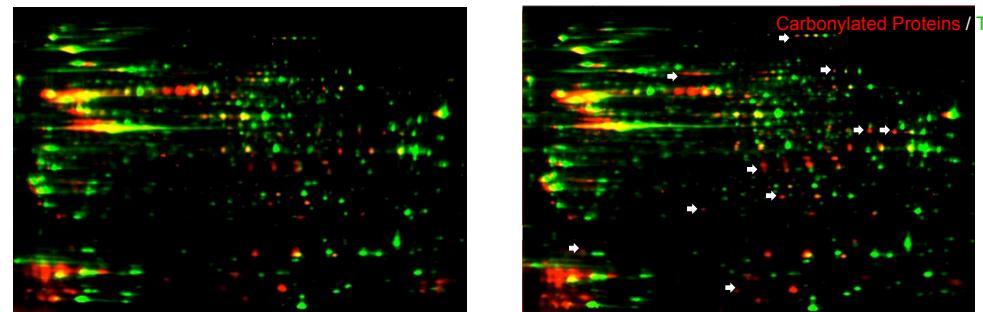
VALIDATION IN BIOLOGICAL SAMPLES FROM DIFFERENT ORIGINS

Seminal Plasma and infertility



Faure et al. 2014, PLoS One

Mitochondrial oxidative stress in cancer cells



Rodier et al. 2015, Cell Reports



Martin BARAIBAR
Janek HYZEWICZ
Emad AHMED*
Liang LIU*
Sofia LOURENCO DOS SANTOS
Audrey DESVERGNE
Marie-Paule HAMON
Sabrina RADJEI*
Romain LADOUCE*
Marine LE BOULCH*
Monique GAREIL
Hilaire BAKALA
Isabelle PETROPOULOS

Alexander BÜRKLE & Maria MORENO-VILLANUEVA
University of Konstanz, Germany

Adelina ROGOWSKA-WRZESINSKA & Peter ROEPSTORFF
University of Odense, Denmark.

Gillian BUTLER-BROWNE & Vincent MOULY
INSERM-UPMC, Paris, France.

Carina PRIP-BUUS & Anne-Laure BULTEAU
Institut Cochin, INSERM-CNRS, Paris, France

Brian CLARK
Peter KRISTENSEN
University of Aarhus
Denmark

Lars LARSSON
Karolinska Institute
Stockholm, Sweden

Tilman GRUNE
DIFE, Nuthetal,
Germany



EU/FP6 - IP 518230 (2006-11)

EU/FP7 - IP 200880 (2008-14)



EU/FP7 - IP 223576 (2009-14)



AFM (2013-16)



EU/FP7 – COST CM 1001 (2010-14)