

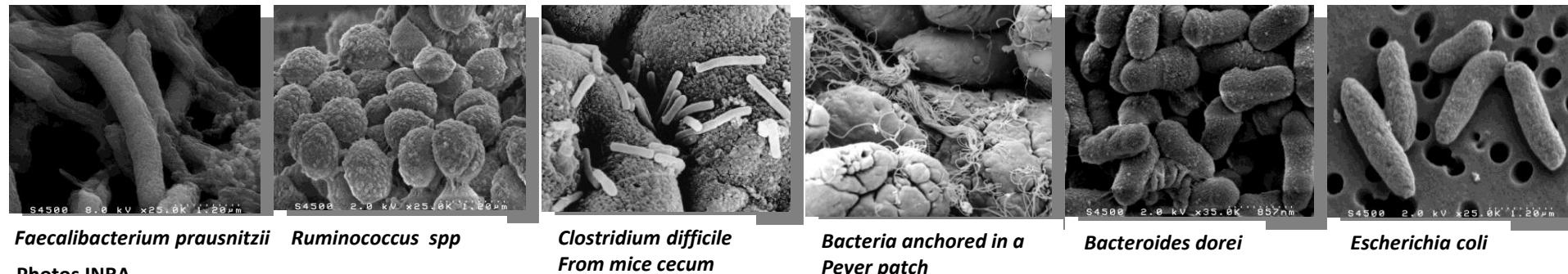
# INTERACTIONS MICROBIOTE/HÔTE APPROCHES FONCTIONNELLES

**Hervé M. Blottière**

FInE lab « Functionality of the Intestinal Ecosystem »,  
Micalis Institute, INRA, AgroParisTech, Université Paris-Saclay  
&  
MetaGenoPolis, INRA, Université Paris-Saclay  
78350 Jouy en Josas

# THE HUMAN INTESTINAL MICROBIOTA

- ✓ 39 trillions microorganisms (Sender et al, Cell 2016) ;
- ✓ As many microorganisms as human cells
- ✓ Hundreds of different species predominantly **not yet cultured** (~70% of dominant species);
- ✓ a few dozen species conserved between individuals (core); a stable community.
- ✓ A key organ, interacting with food (fermentation,...); interacting with our cells (immune & nervous systems,...); protecting against pathogens (barrier function);...
- ✓ A true organ, revealed as playing a role in several diseases
- ✓ Thousands of metabolites/molecules with potential interest (Blotti  re & Dor  , m  decine/science, 2016)



# METAGENOME : genomes of all dominant microbes in an ecosystem

## A revolution since the turn of the century



Bacterial fraction

14/16 - 1 -

### Lessons from early human intestinal tract metagenomics:

- Qin *Nature* 2010 => 3.3 million gene catalog ; both core metagenome & rare genes
- Arumugam *Nature* 2011 => 3 enterotypes, preferred ecological arrangements
- Schloissnig *Nature* 2012 => stability at SNPs level (strains)
- Qin *Nature* 2012 => metagenomic dysbiosis in T2D
- Le Chatelier *Nature* 2013 => metagenomic dysbiosis in obesity ; diagnostic signatures
- Cotillard *Nature* 2013 => metagenomic dysbiosis in obesity ; low gene count as stratifier
- Qin *Nature* 2014 => metagenomic dysbiosis in liver cirrhosis
- Li *Nat Biotech* 2014 => 10 million gene catalog ; core metagenome unchanged
- Nielsen *Nat Biotech* 2014 => co-abundant gene clustering and metagenomic species
- Xiao *Nat Biotech* 2015 => mouse gene catalog ; environment dependent
- Shoae *Cell Metabol* 2015 => nutrition and intestinal metabolome
- Forslund *Nature* 2015 => metformin signature in T2D
- Xiao *Nature Microbiol* 2015 => A pork gut reference catalog
- Dao *Gut* 2016 => Akkermansia and obesity/MetS
- Plichta *Nature Microbiol* 2016 => metatranscriptomics, niche segregation
- Pedersen *Nature* 2016 => microbiota affect insulin sensitivity
- Costea, *Nat Biotech*, 2017 => SOPs
- Routy, *Science*, 2017 => Microbiota and success of cancer therapy



How to go further and introduce  
microbiota in human health ?

# MetaGenoPolis



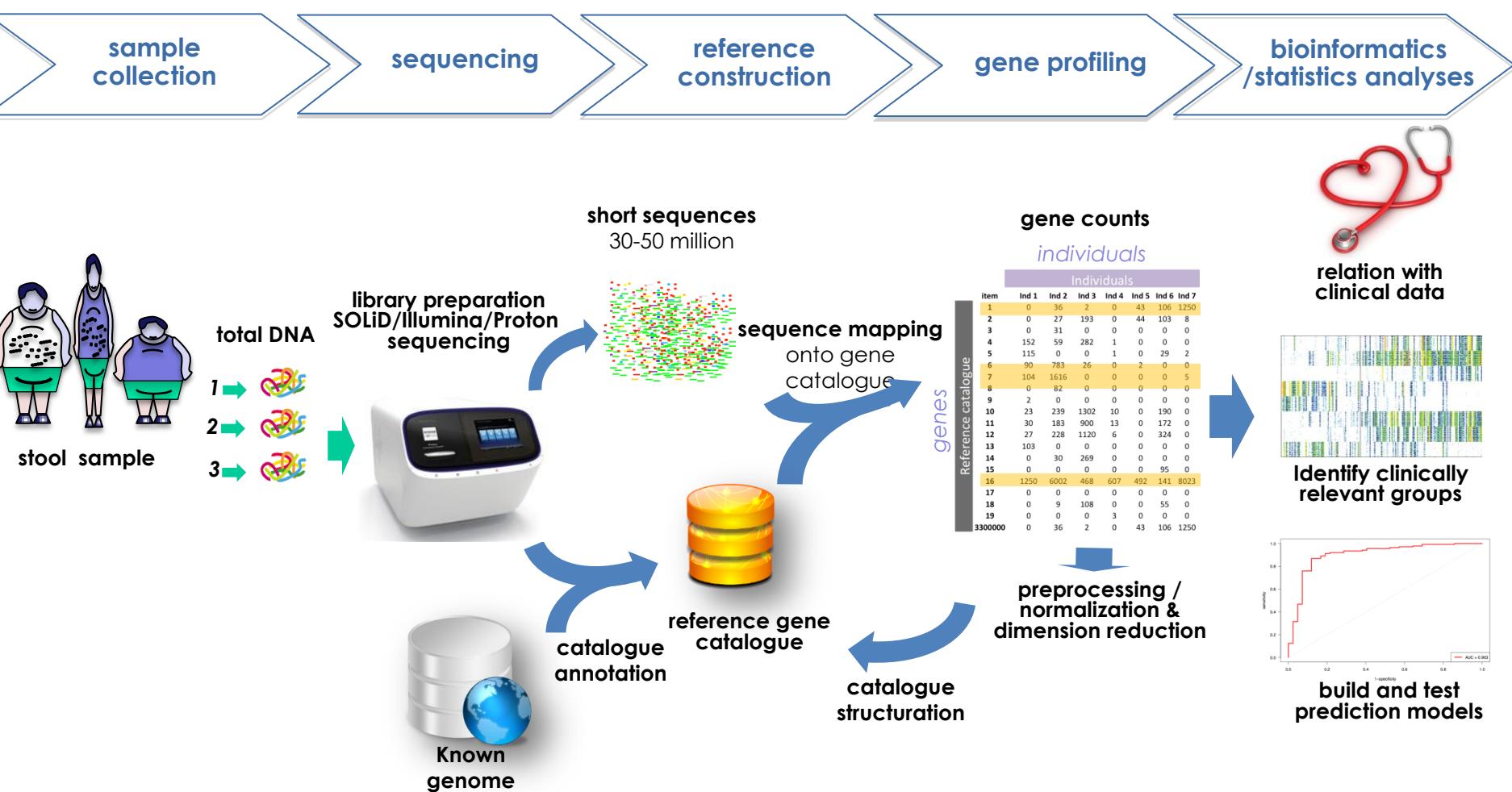
Platforms dedicated to  
quantitative and functional  
metagenomics



*Investment for the Future,  
FRANCE*

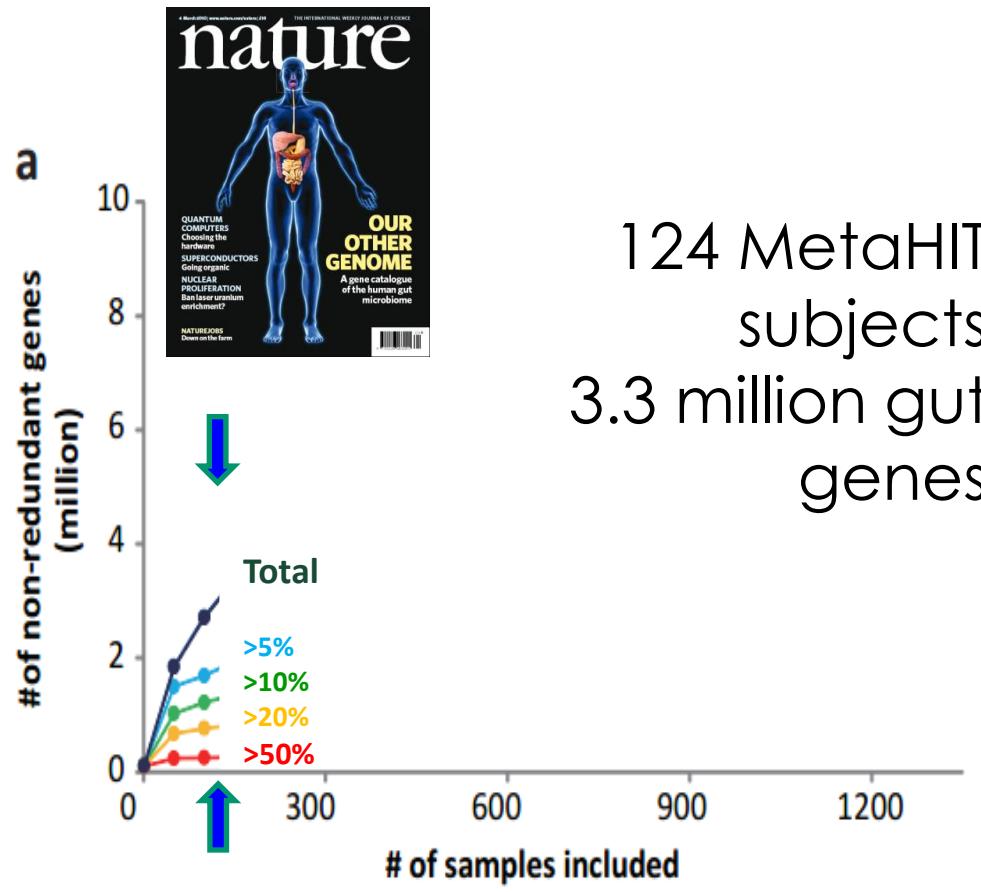
# QUANTITATIVE METAGENOMIC PIPELINE AT METAGENOPOLIS

## SAMBO-METAQUANT-INFOBIOSTAT



4000 samples/yr

# A REFERENCE CATALOG OF 3.3 MILLION GUT GENES



124 MetaHIT  
subjects  
3.3 million gut  
genes

Rare genes are  
increasing



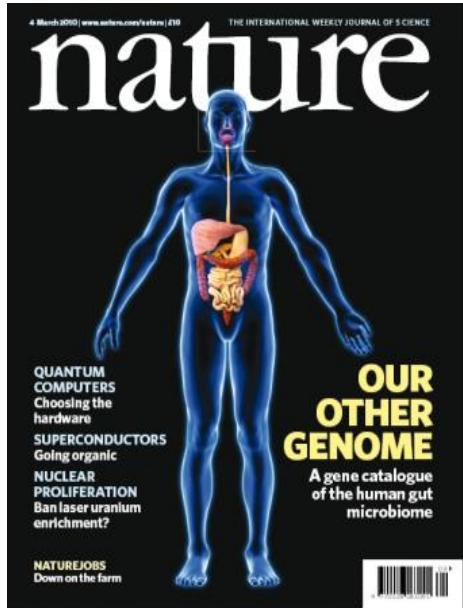
Common genes are not

Individuals from MetaHIT, Chinese and HMP studies, n=1267

Qin, *Nature* 2010

Li et al. *Nature Biotechnol.*, 2014

# A REFERENCE CATALOG OF 10 MILLION GUT GENES



124 MetaHIT EU subjects  
3.3 million gut genes



Individuals from MetaHIT, Chinese  
and HMP studies, n=1267

On average, each individual carries ~540 000 genes  
of the initial catalog. 50 % of the genes of an individual are shared  
by at least 50 % of individuals = **core metagenome**

Qin, Nature 2010

Li et al. Nature Biotechnol, 2014

# PERTURBATION OF INTESTINAL MICROBIOTA AS A POSSIBLE CHRONIC DISEASE FACTOR

also

- ✓ Multiple sclerosis
- ✓ Alzheimer disease
- ✓ Parkinson disease
  
- ✓ Hypertension
- ✓ Bone
- ✓ Kidney diseases
- ✓ ...

Human clinical studies :  
**Shotgun sequencing**  
16S sequencing

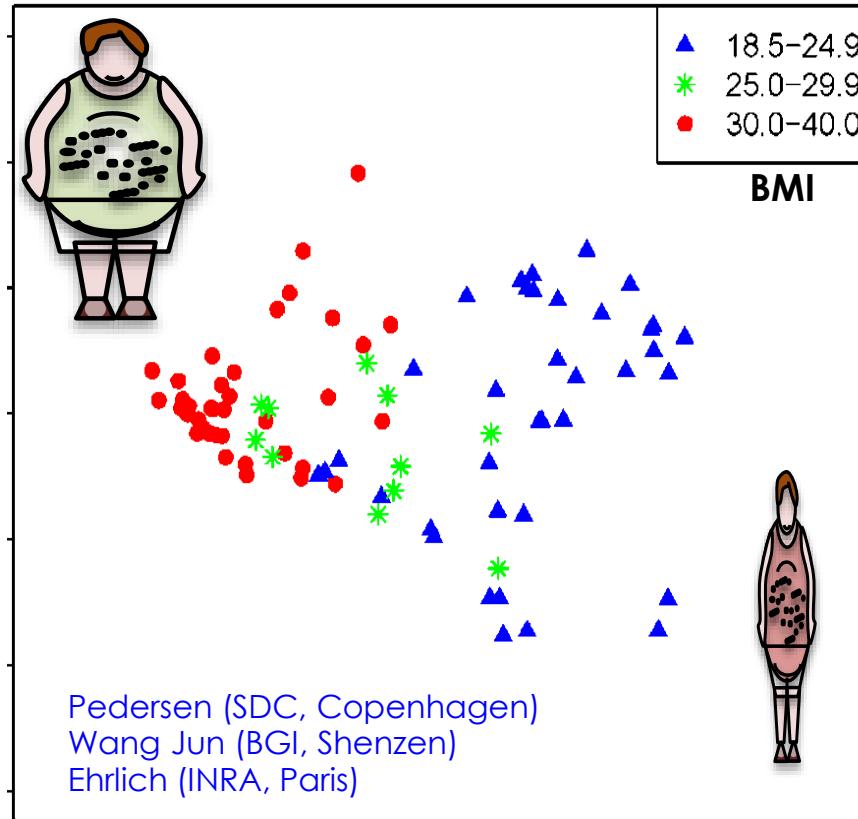
Blottière & Doré,  
médecine/science, 2016

Pathologies	References
<b>Crohn's disease</b>	<b>Qin, Nature 2010</b> Gevers, Cell Host Microbe 2014
<b>Ucerative colitis</b>	<b>Qin, Nature 2010</b> Lepage , Gastroenterology, 2011
<b>Celiac disease</b>	D'Argenio, Am J Gastroenterol 2016
<b>Irritable bowel syndrome</b>	Saulnier, Gastroenterology 2011 Rajilic-Stojanovic, Gastroenterology 2011
<b>Colorectal cancer</b>	<b>Zeller, Mol Syst Biol 2014</b> Sobhani PLoS one 2011
<b>Obesity</b>	<b>Le Chatelier, Nature 2013</b> Ley, Nature 2006
<b>Type 1 diabetes</b>	<b>Kostic, Cell Host Microbes 2015</b> Murri, BMC medicine 2012
<b>Type 2 diabetes</b>	<b>Forslund, Nature 2015</b>
<b>Seniors frailty</b>	Claesson Nature 2012
<b>GVHD</b>	Taur, Blood, 2014
<b>Allergy</b>	Abrahamsson, J Allergy Clin Immunol 2012
<b>Liver pathologies</b>	<b>Qin, Nature 2014</b>
<b>Cardiovascular diseases</b>	Karlsson Nat Commun 2012 Projet MetaCardis
<b>Autism - Depression</b>	Finegold, Angerobe 2010

# METAGENOMIC SIGNATURES OF DYSBIOSIS IN OBESITY



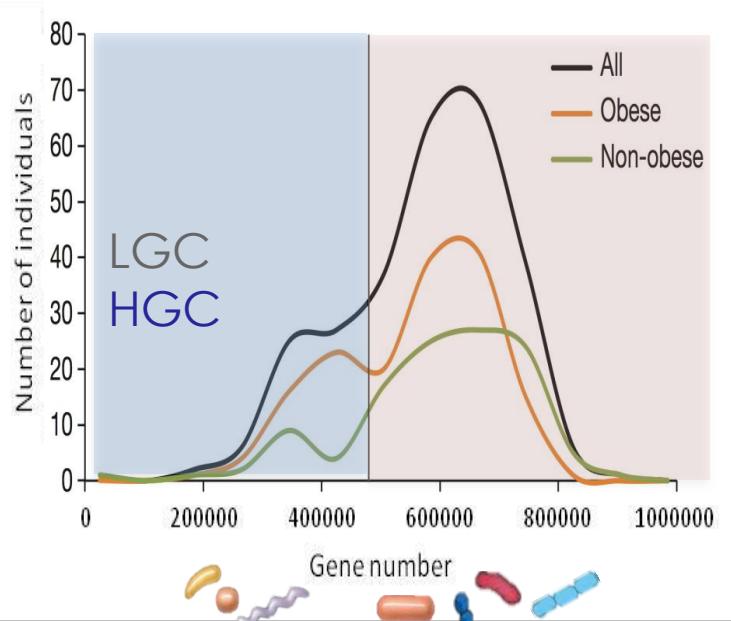
Animal studies demonstrated a link between obesity and gut microbiota



Bacterial genes and genomes specific of the microbiome of patients

# MICROBIOTA GENE COUNT / DIVERSITY IS A HEALTH-ASSOCIATED STRATIFIER

Low to High  
gene count (French or Danes)



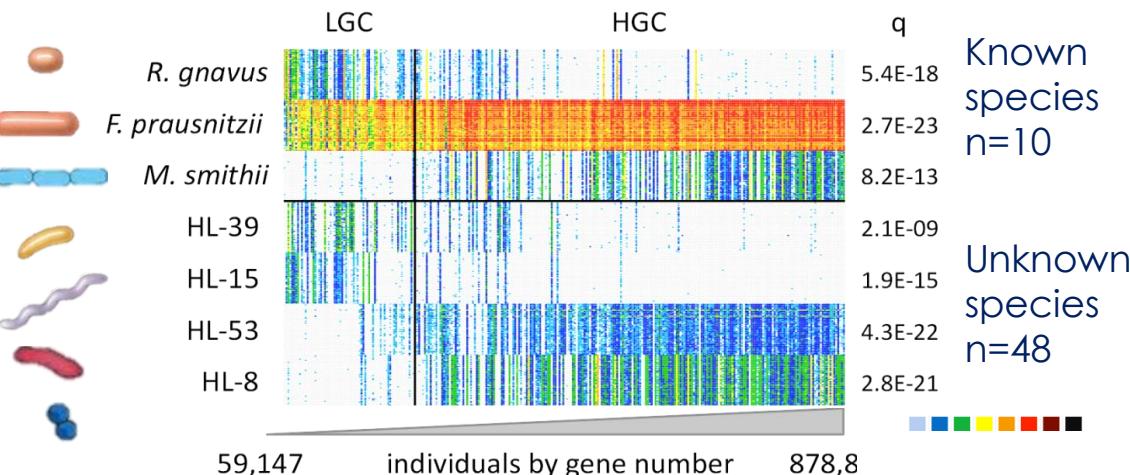
**LGC associates with CMD risks**

- ↑↑ dyslipidemia
- ↑ adiposity
- ↑ insulin resistance
- ↑ inflammation (circulating and adipose tissue)

and healthier diet



## Signature species (n= 58)



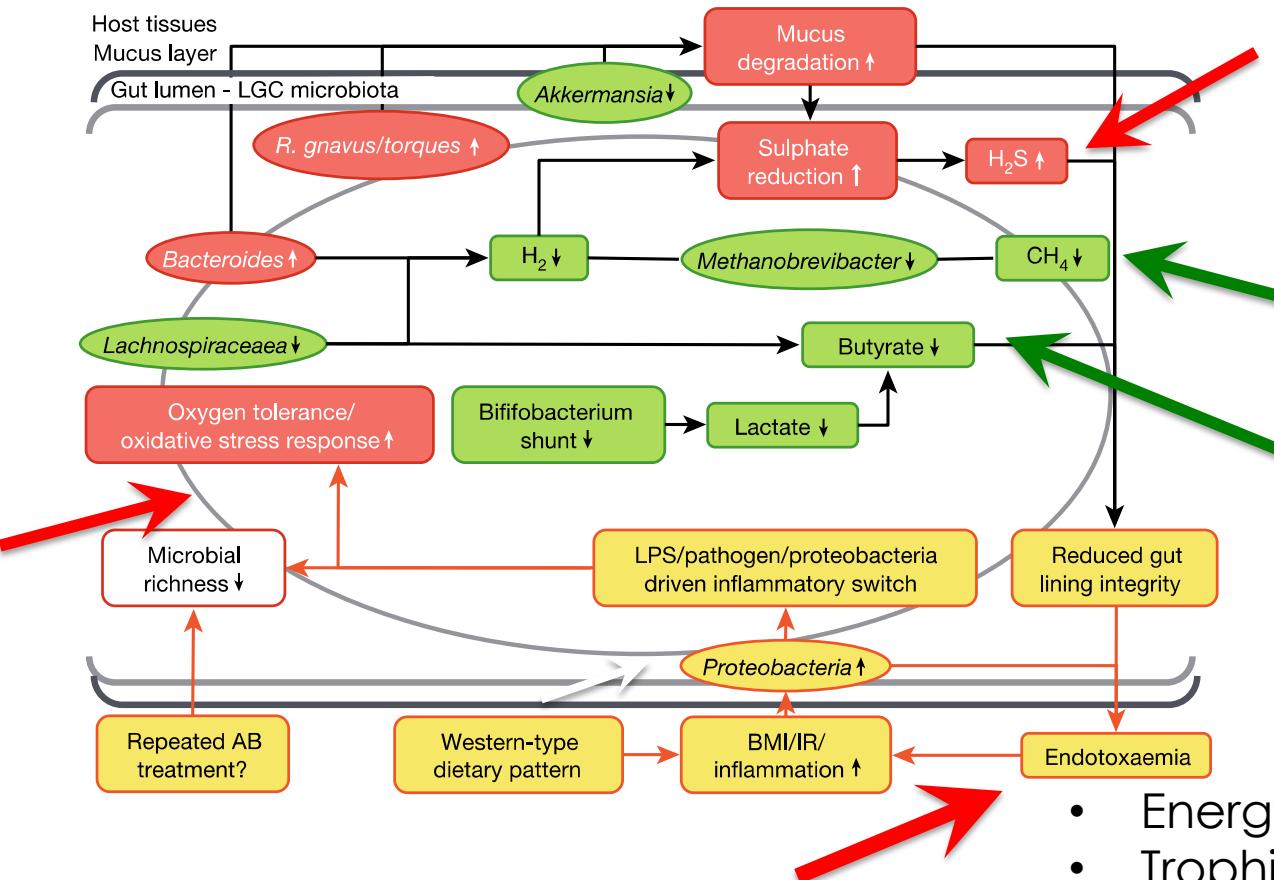
LGC: ↑ Pro-inflammatory

HGC: ↑ Anti-inflammatory

- Cl. bolteae*
- Cl. symbiosum*
- Cl. clostridioforme*
- Cl. ramosum*
- R. gnavus*
- F. prausnitzii*
- R. inulinivorans*
- Co. eutactus*
- M. smithii*



# FUNCTIONAL SHIFTS IN THE LGC MICROBIOME



- Energy source for IEC
- Trophic & barrier functions
- HDAC inhibitor (gene regulation)
- GPR agonist – PYY & GLP1
- Immunomodulatory effects

Le chatelier et al, Nature, 2013

Blottière et al, PNS 2003;  
Segain et al, Gut, 2000

# LETTER

doi:10.1038/nature12721



## Commensal microbe-derived butyrate induces the differentiation of colonic regulatory T cells

Yukihiro Furusawa<sup>1,2\*</sup>, Yuuki Obata<sup>1,2,3\*</sup>, Shinji Fukuda<sup>1,4\*</sup>, Takaho A. Endo<sup>1</sup>, Gaku Nakato<sup>1</sup>, Daisuke Takahashi<sup>1</sup>, Yumiko Nakanishi<sup>4</sup>, Chikako Uetake<sup>1</sup>, Keiko Kato<sup>1,5</sup>, Tamotsu Kato<sup>1</sup>, Masumi Takahashi<sup>1</sup>, Noriko N. Fukuda<sup>4</sup>, Shinnosuke Murakami<sup>4</sup>, Eiji Miyauchi<sup>1</sup>, Shingo Hino<sup>6</sup>, Koji Atarashi<sup>1,7</sup>, Satoshi Onawa<sup>1</sup>, Yumiko Fujimura<sup>2</sup>, Trevor Lockett<sup>8</sup>, Julie M. Clarke<sup>8</sup>, David L. Topping<sup>8</sup>, Masaru Tomita<sup>4</sup>, Shohei Horii<sup>1</sup>, Osamu Ohara<sup>1</sup>, Tatsuya Morita<sup>6</sup>, Haruhiko Koseki<sup>1,3,5</sup>, Jun Kikuchi<sup>5,9</sup>, Kenna Honda<sup>1,10</sup>, Koii Hase<sup>1,2,7\*</sup> & Hiroshi Ohno<sup>1,3,5</sup>

# LETTER

doi:10.1038/nature12726

## Metabolites produced by commensal bacteria promote peripheral regulatory T-cell generation

Nicholas Arpaia<sup>1,2</sup>, Clarissa Campbell<sup>1,2</sup>, Xiying Fan<sup>1,2</sup>, Stanislav Dikiy<sup>1,2</sup>, Joris van der Veeken<sup>1,2</sup>, Paul deRoos<sup>1,2</sup>, Hui Liu<sup>3</sup>, Justin R. Cross<sup>3</sup>, Klaus Pfeffer<sup>4</sup>, Paul J. Coffer<sup>1,2,5</sup> & Alexander Y. Rudensky<sup>1,2</sup>

19 / 26 DECEMBER 2013 | VOL 504 | NATURE | 451

Cell  
PRESS

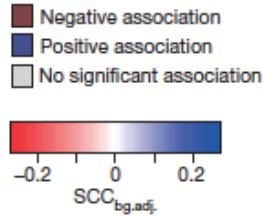
Immunity  
Article

## Activation of Gpr109a, Receptor for Niacin and the Commensal Metabolite Butyrate, Suppresses Colonic Inflammation and Carcinogenesis

Nagendra Singh,<sup>1,2,\*</sup> Ashish Gurav,<sup>1</sup> Sathish Sivaprakasam,<sup>1</sup> Evan Brady,<sup>1</sup> Ravi Padia,<sup>1</sup> Huidong Shi,<sup>1,2</sup> Muthusamy Thangaraju,<sup>1,2</sup> Puttur D. Prasad,<sup>1,2</sup> Santhakumar Manicassamy,<sup>2</sup> David H. Munn,<sup>2,3</sup> Jeffrey R. Lee,<sup>4</sup> Stefan Offermanns,<sup>5</sup> and Vadivel Ganapathy<sup>1,2,\*</sup>

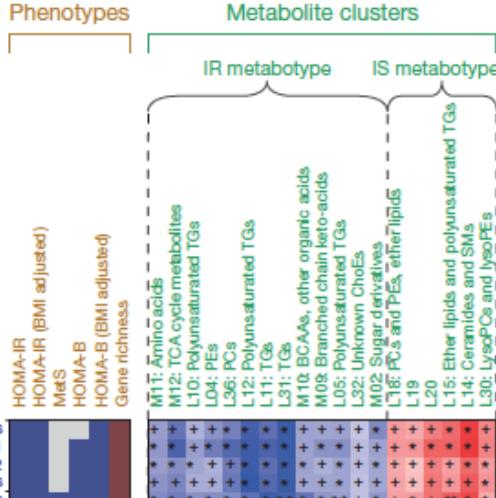
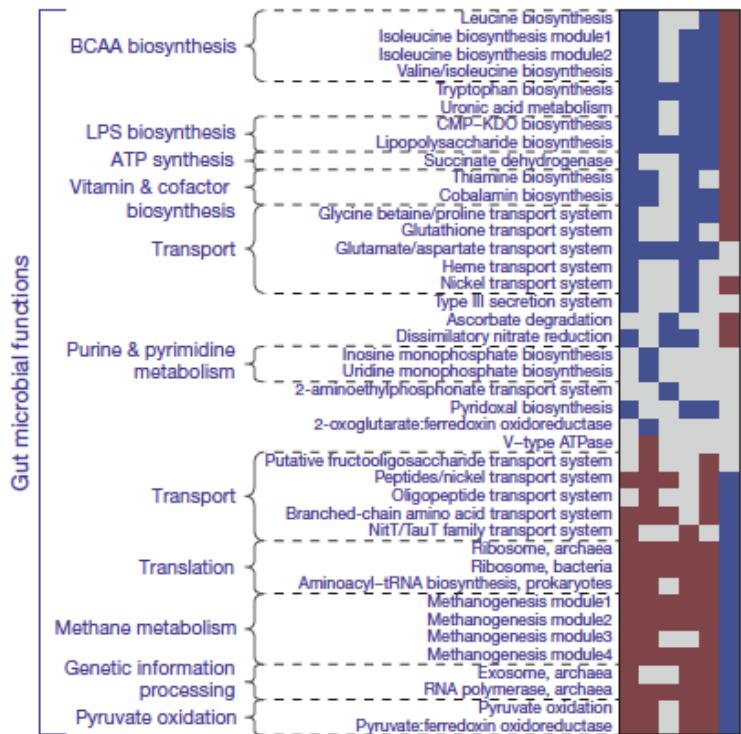
 INRA  
SCIENCE & IMPACT

# GUT MICROBIOME, SERUM METABOLOME & INSULIN RESISTANCE



insulin sensitivity and MetS

## Feces metagenome

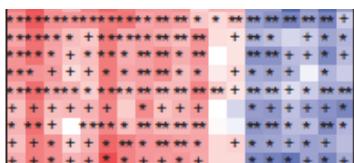


277 non-diabetic individuals  
(164 obese, 113 non-obese)

fasting serum metabolome

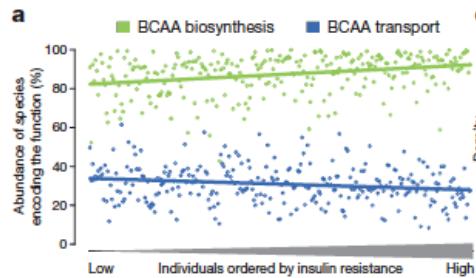
Functions of IR-associated microbiome have an impact on the metabolome :

- LPS and **BCAA** biosynthesis
- **BCAA** transport
- Methanogenesis

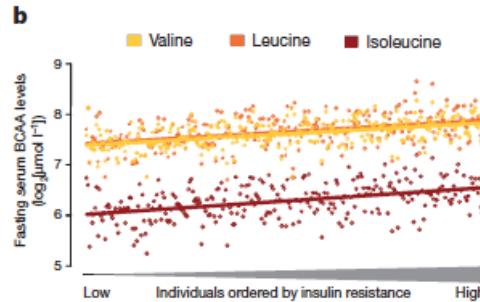


# BRANCH CHAIN AMINO-ACIDS & INSULIN RESISTANCE

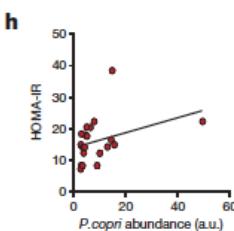
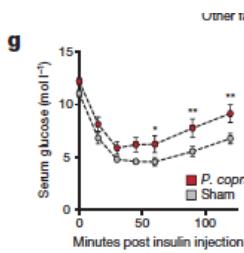
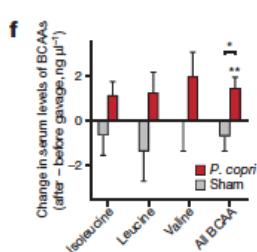
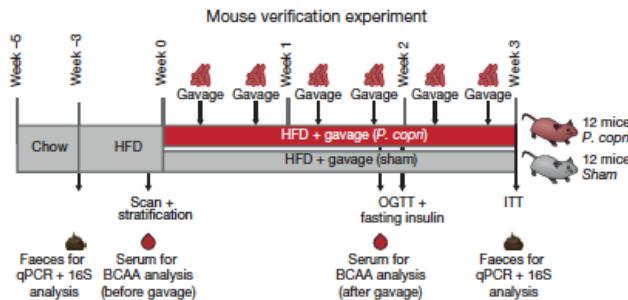
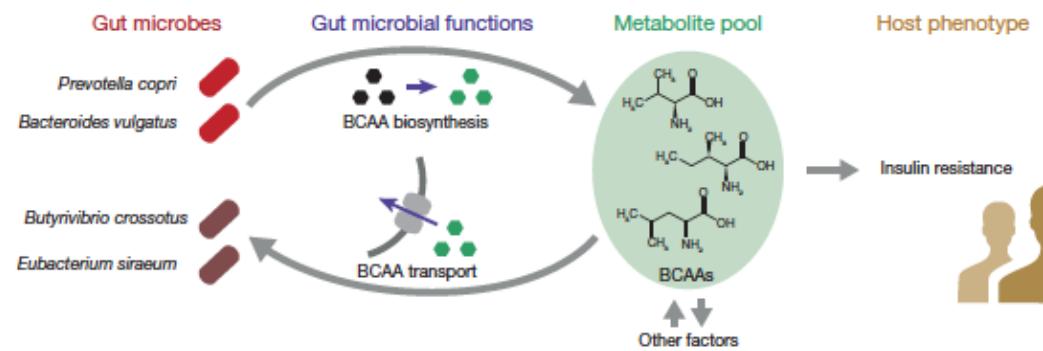
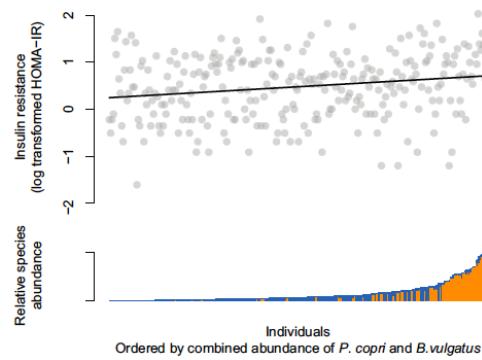
## Microbiome functions



## Metabolome

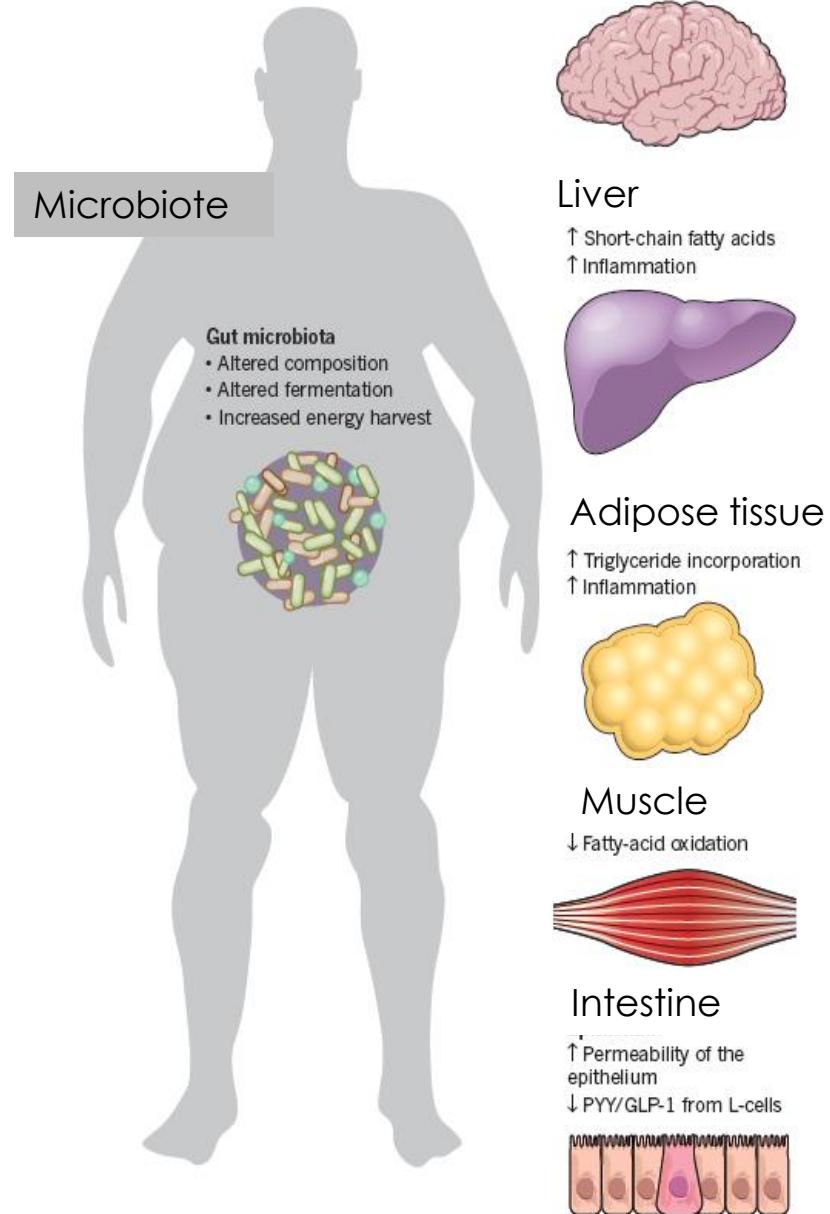
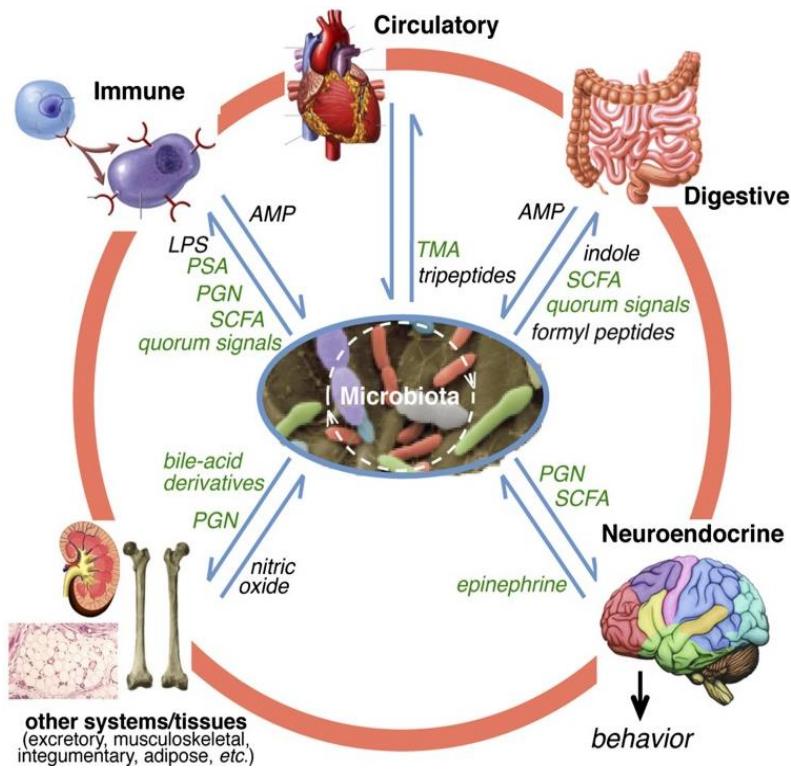


**b**



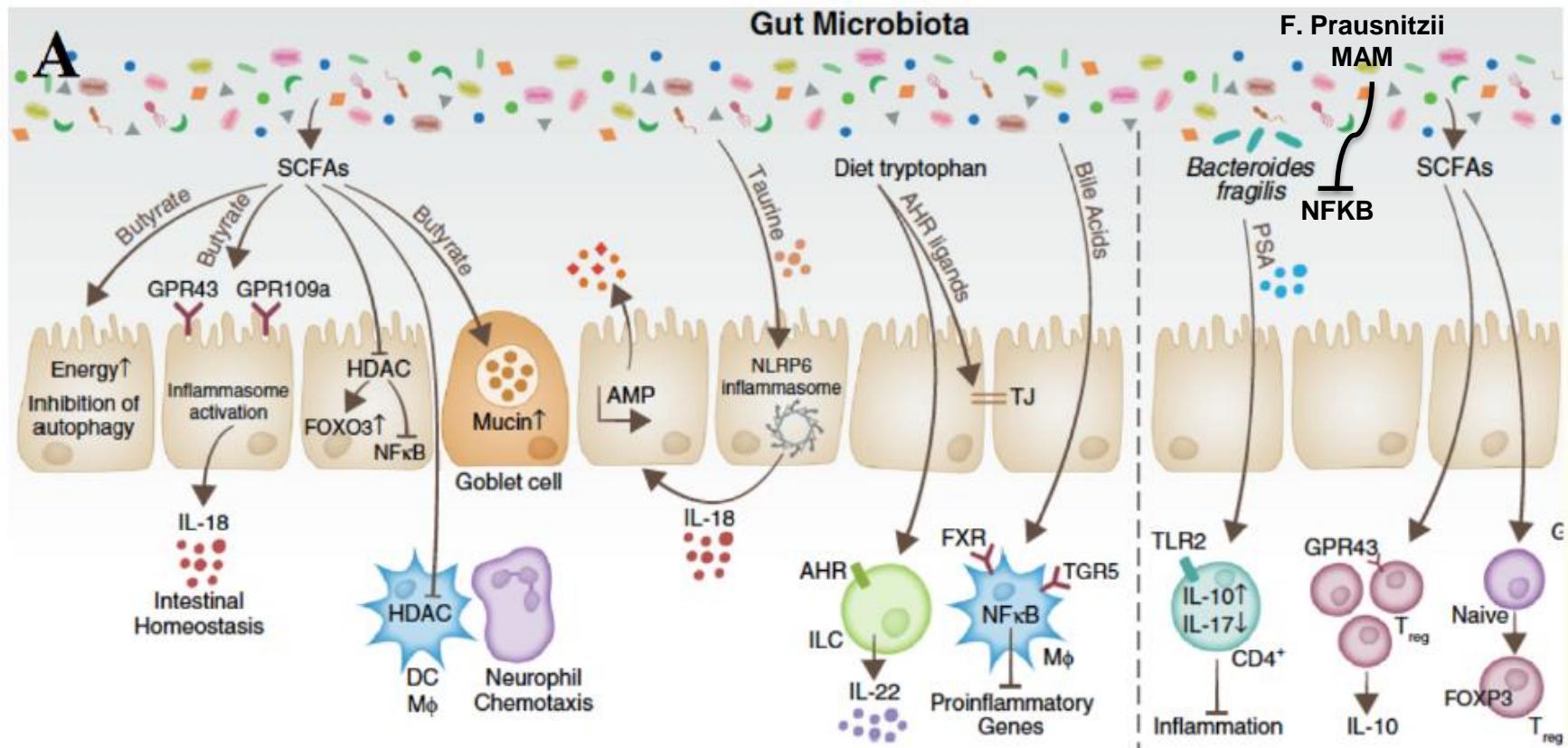
**INRA**  
SCIENCE & IMPACT

# MECHANISMS LINKING DYSBIOSIS AND DISEASES



Tremaroli et Bäckhed, Nature 2012;  
McFall-Ngai et al, 2013

# HOST-MICROBIOTA INTERACTIONS. FEW MOLECULES IDENTIFIED



Ligand for AhR

# MICROBIOTA HOST-INTERACTIONS

## USE OF ANIMAL MODELS

- ✓ The intestinal microbiota plays a key role on Human Health, but how ?
- ✓ Few bacterial metabolites are known to control/modulate host cell response, i.e. SCFA, TMA, indols, ... only few are known
- ✓ Can we learn from animal model ?



# GUT MICROBIOTA AND IMMUNE SYSTEM LESSON FROM GERM-FREE ANIMALS

## GERMFREE LIFE AND GNOTOBIOLOGY

By THOMAS D. LUCKEY



ACADEMIC PRESS · New York · London

1963



Glimstedt G. Bakterienfrei Meerschweinchen. Aufzucht, Lebensfähigkeit und Wachstum, nebst Untersuchung über das lymphatische Gewebe. Acta Pathol Microbiol Scand. 1936;S30:1–295.

Bacteria free guinea pigs. Rearing, viability and growth, as well as investigation of the lymphatic tissue.

# MAIN IMMUNE SYSTEM ABNORMALITIES DESCRIBED IN GERM-FREE MICE COMPARED TO CONVENTIONAL ANIMALS



Immunological deficiency	Site	Phenotype in axenic mice
Development of the small intestine	Peyer's patches	Decrease in number and size
	Lamina propria	Thinner, fewer lymphocytes
	Germinal centers	Fewer plasmacytoid dendritic cells
	Isolated lymphoid follicles	Smaller
Development of mesenteric lymph nodes	Germinal centers	Smaller with fewer plasmacytoid dendritic cells
CD8+ T cells	Intraepithelial lymphocytes	Less numerous and less cytotoxic
CD4+ T cells	Lamina propria	Reduced cell number, decrease in Th17 in the small intestine but not in the colon
MAIT cells		Absent
CD4+ and CD8+ T cells	Spleen	Reduced cell number, reduced cytokine production
CD4+CD25+ T cells	Mesenteric lymph nodes	Decreased expression of FoxP3 and reduced suppressive capacities
Expression of antimicrobial peptides	Paneth cells	Reduced
IgA production	B cells	Reduced, and fewer serum IgA
IgG and IgM	Serum	Strongly reduced levels
ATP level	Gut	Reduced
Expression of class II MHC	Intestinal epithelial cells	Reduced
Expression of TLR2, TLR4 and TLR9		
IL-25 level		Reduced

# THE INNATE IMMUNE SYSTEM AND GUT MICROBIOTA

Cell 145, 745–757, May 27, 2011 ©2011 Elsevier Inc. 745

Cell

## NLRP6 Inflammasome Regulates Colonic Microbial Ecology and Risk for Colitis



Eran Elinav,<sup>1,8</sup> Till Strowig,<sup>1,8</sup> Andrew L. Kau,<sup>4,5</sup> Jorge Henao-Mejia,<sup>1</sup> Christoph A. Thaiss,<sup>1</sup> Carmen J. Booth,<sup>2</sup> David R. Peaper,<sup>3</sup> John Bertin,<sup>6</sup> Stephanie C. Eisenbarth,<sup>1,3</sup> Jeffrey I. Gordon,<sup>4</sup> and Richard A. Flavell<sup>1,7,\*</sup>

Cell

1428 Cell 163, 1428–1443, December 3, 2015 ©2015 Elsevier Inc.

Article

## Microbiota-Modulated Metabolites Shape the Intestinal Microenvironment by Regulating NLRP6 Inflammasome Signaling

Maayan Levy,<sup>1,13</sup> Christoph A. Thaiss,<sup>1,13</sup> David Zeevi,<sup>2,3</sup> Lenka Dohnalová,<sup>1</sup> Gilli Zilberman-Schapira,<sup>1</sup> Jemal Ali Mahdi,<sup>1,4</sup> Eyal David,<sup>1</sup> Alon Savidor,<sup>5</sup> Tal Korem,<sup>2,3</sup> Yonatan Herzig,<sup>1</sup> Meirav Pevsner-Fischer,<sup>1</sup> Hagit Shapiro,<sup>1</sup> Anette Christ,<sup>6,7</sup> Alon Hammelin,<sup>8</sup> Zamir Halpern,<sup>9,10</sup> Eicke Latz,<sup>6,7</sup> Richard A. Flavell,<sup>11,12</sup> Ido Amit,<sup>1</sup> Eran Segal,<sup>2,3,14,\*</sup> and Eran Elinav<sup>1,14,\*</sup>

# CAUTION WITH ANIMAL MODEL

Please cite this article in press as: Mamantopoulos et al., Nlrp6- and ASC-Dependent Inflammasomes Do Not Shape the Commensal Gut Microbiota Composition, *Immunity* (2017), <http://dx.doi.org/10.1016/j.immuni.2017.07.011>

Immunity

Article

## Nlrp6- and ASC-Dependent Inflammasomes Do Not Shape the Commensal Gut Microbiota Composition

Michail Mamantopoulos,<sup>1,2,12</sup> Francesca Ronchi,<sup>3,12</sup> Filip Van Hauwermeiren,<sup>1</sup> Liesbet Martens,<sup>2,6</sup> Yvan Saeys,<sup>2,7</sup> Stefan K. Drexler,<sup>8</sup> Amir S. Yazdi,<sup>9</sup> Jeanne K. McCoy,<sup>3,10,11,13,\*</sup> and Andy Wullaert<sup>1,2,11,\*</sup>

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<sup>4</sup>Department of Microbiology and Immunology, KU Leuven, Rega Institute, Leuven, Belgium

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<sup>7</sup>Department of Applied Mathematics, Computer Science and Statistics, Ghent University, Ghent, Belgium

<sup>8</sup>Biozentrum, University of Basel, 4056 Basel, Switzerland

<sup>9</sup>Department of Dermatology, University of Tübingen, Tübingen, Germany

<sup>10</sup>Department of Physiology and Pharmacology and Calvin, Phoebe and Joan Snyder Faculty of Veterinary Medicine, University of Calgary, Calgary, AB, Canada

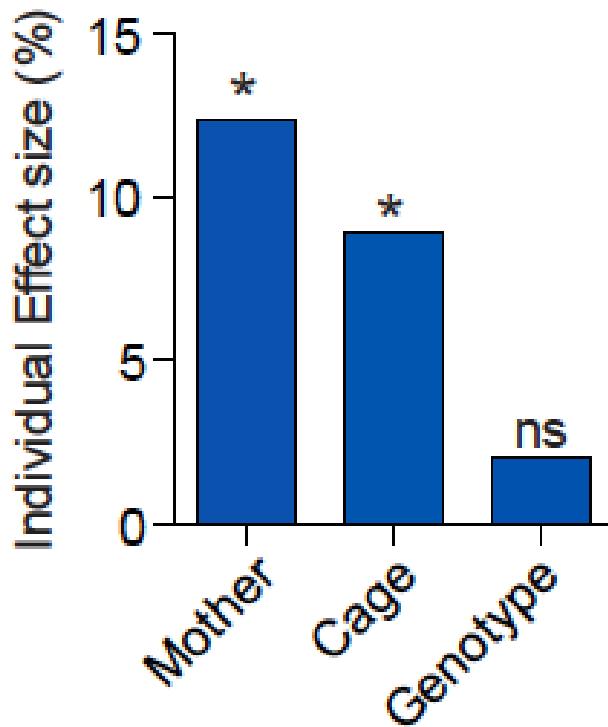
<sup>11</sup>Senior author

<sup>12</sup>These authors contributed equally

<sup>13</sup>Lead Contact

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F



# CAUTION WITH ANIMAL MODEL

Please cite this article in press as: Rosshart et al., Wild Mouse Gut Microbiota Promotes Host Fitness and Improves Disease Resistance, *Cell* (2017), <https://doi.org/10.1016/j.cell.2017.09.016>

## Article

Cell

# Wild Mouse Gut Microbiota Promotes Host Fitness and Improves Disease Resistance

Stephan P. Rosshart,<sup>1,\*</sup> Brian G. Vassallo,<sup>1</sup> Davide Angeletti,<sup>1</sup> Kazuyo Takeda,<sup>5</sup> Heather D. Hickman,<sup>2</sup> John A. McCullough,<sup>3</sup> Fernando Pardo-Manuel de Villena,<sup>4</sup> Jonathan W. Yewdell,<sup>6</sup>

<sup>1</sup>Immunology Section, Diseases Branch, National Institute of Diabetes and Digestive and Kidney Diseases (NIDDK), National Institutes of Health (NIH), Bethesda, MD 20892, USA

<sup>2</sup>Laboratory of Viral Diseases, National Institute of Allergy and Infectious Diseases (NIAID), NIH, Bethesda, MD 20892, USA

<sup>3</sup>Alkek Center for Metagenomics and Microbiome Research, Department of Molecular Biology and Biochemistry, University of Texas at Houston, TX 77030, USA

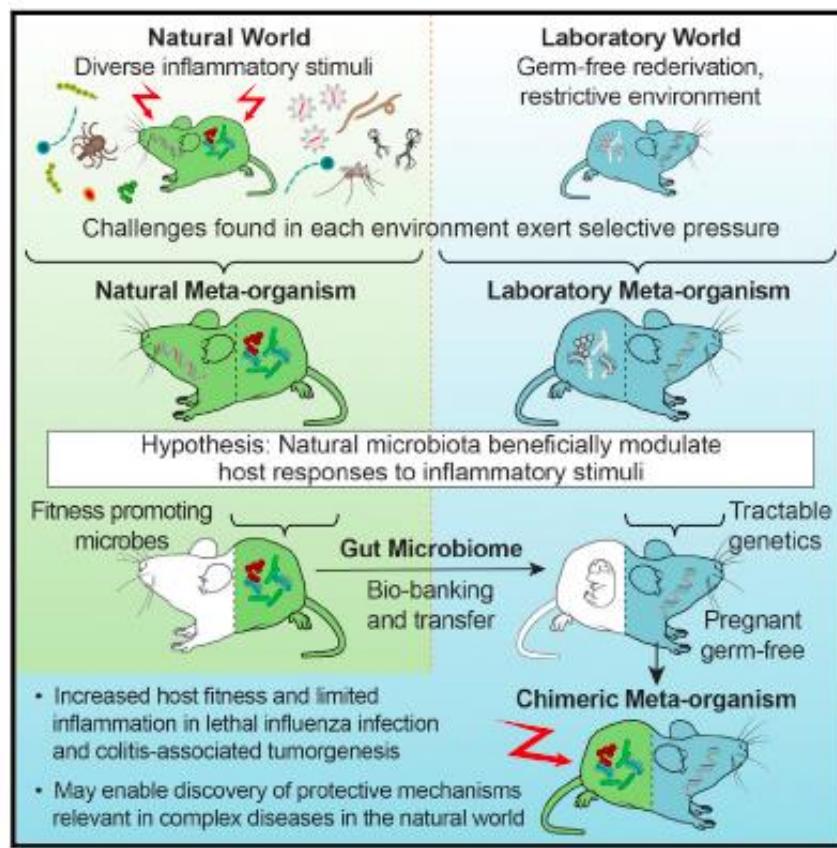
<sup>4</sup>Department of Genetics, Lineberger Comprehensive Cancer Center, University of North Carolina at Chapel Hill, Chapel Hill, NC 27599, USA

<sup>5</sup>Microscopy and Imaging Core Facility, Center for Biologics Evaluation and Research, NIH, Bethesda, MD 20893-0002, USA

<sup>6</sup>Cancer and Inflammation Program, Center for Cancer Research, NIH, Bethesda, MD 20892, USA

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\*Correspondence: [stephanpatrick.rosshart@nih.gov](mailto:stephanpatrick.rosshart@nih.gov) (S.P.R.), [rehermann@nih.gov](mailto:rehermann@nih.gov) (B.G.V.)



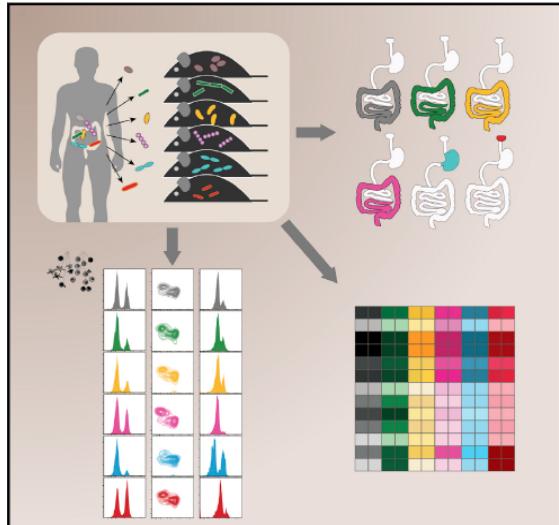
WORK WITH CHIMERIC MICE

# INTERACTION WITH IMMUNE SYSTEM IS SPECIES SPECIFIC ?

Cell

## Mining the Human Gut Microbiota for Immunomodulatory Organisms

### Graphical Abstract



Resource

### Authors

Naama Geva-Zatorsky, Esen Sefik,  
Lindsay Kua, ..., Diane Mathis,  
Christophe Benoist, Dennis L. Kasper

### Correspondence

dennis\_kasper@hms.harvard.edu

### In Brief

Each of 53 human-resident bacterial species studied in monoculture in mice modulates the host immune system, providing a baseline for investigating how consortia of gut microbes interact with their host.

### Highlights

- Human gut microbiota comprises a treasure trove of immunomodulatory bacteria
- Diverse and redundant immune and transcriptional responses follow monocolonization
- Immunologic and transcriptional changes are not related to microbial phylogeny
- Following monocolonization, immune recalibration varies to strains within a species

Mediators ?

Mechanisms ?

Cell, february 2017

# MICROBIOTA HOST-INTERACTIONS



- ✓ The intestinal microbiota plays a key role on Human Health, but how?
- ✓ Few bacterial response,
- ✓ Animal models
- ✓ How to identify and predict the power of the co-inhabitants of the microbiome



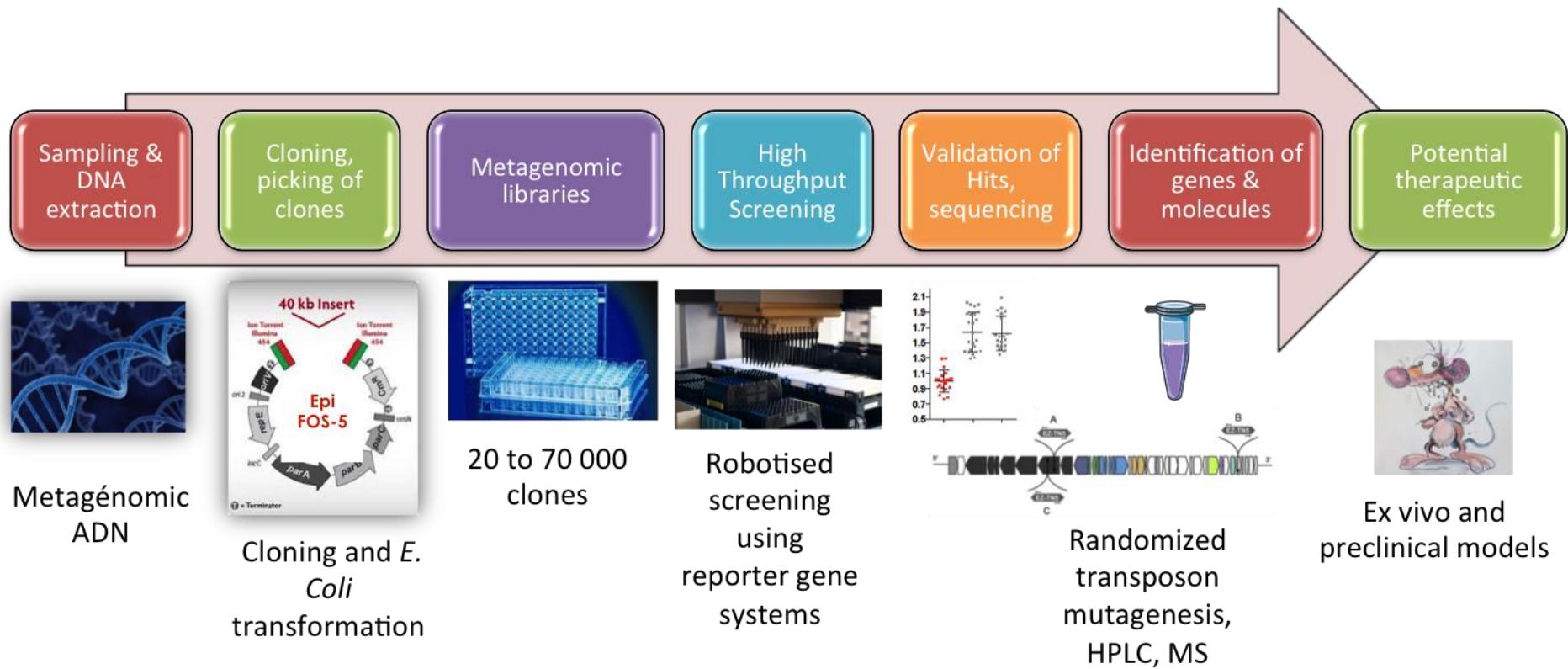
## Functional Metagenomics



to host cell  
k

onal interest  
when 80 %  
stool/tissue

# FUNCTIONAL METAGENOMICS TO STUDY MICROBIOTA – HOST CROSSTALK



Gloux et al., AEM, 2007

Lakhdari et al., Plos one, 2010; de Wouters et al., Plos one, 2014

O'Cuv et al, anaerobes, 2017, 6 patents

From Blottière & Doré, m/s médecin/Sciences, 2016

# FUNCTIONAL METAGENOMICS STRATEGY

Sampling &  
DNA  
extraction

Cloning,  
recombinants  
selection

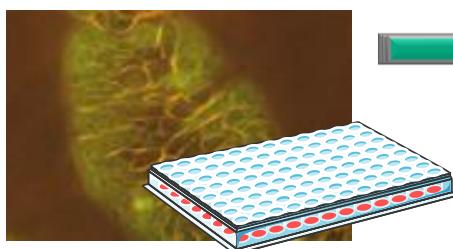
Homogenous  
DNA library

Screening

## Metagenomic libraries



Identification of  
• Clones  
• Genes  
• Molecules



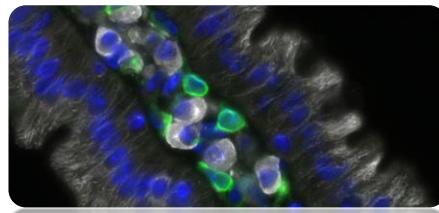
18 libraries  
From healthy,  
obese, Crohn's,  
CRC,  
Ileum/ mucosal  
= 575 000 clones

→ 23 millions  
genes

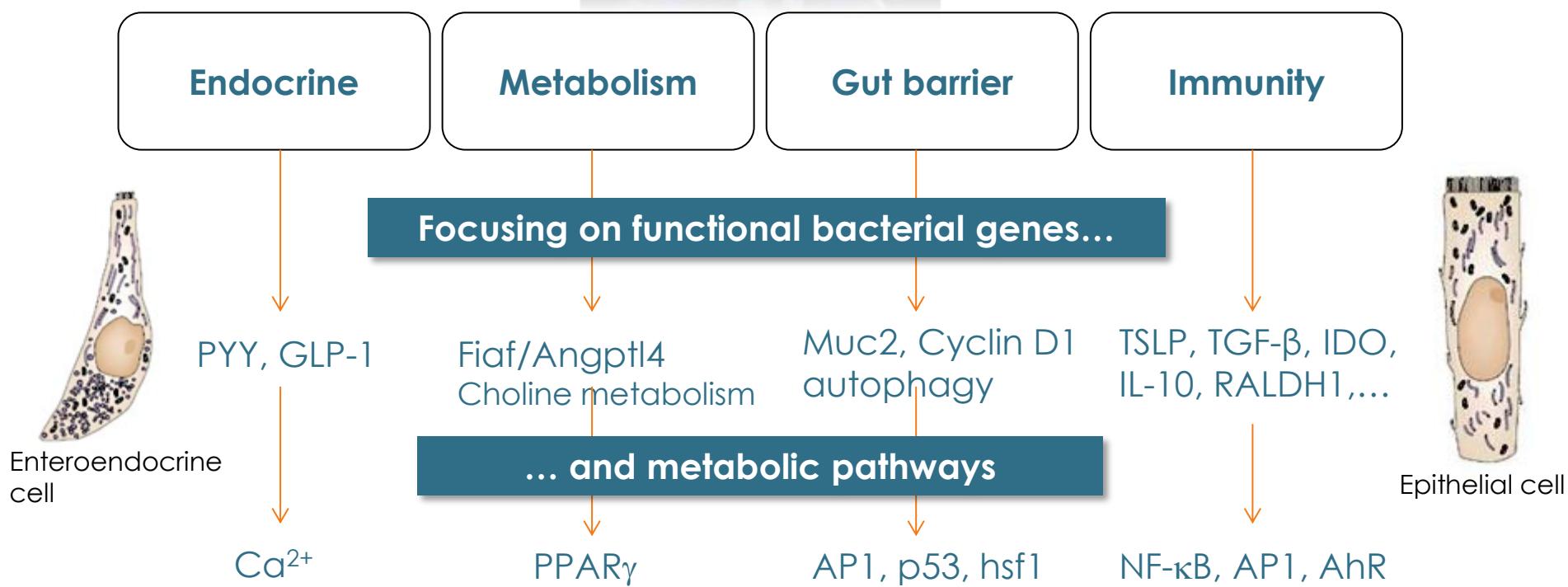
## Reporter human intestinal epithelial cells

Luciferase or fluorescent probes/genes

# TARGETED INTESTINAL FUNCTIONS



Functions linked to several diseases



# ROBOTIC EQUIPMENTS FOR HTS



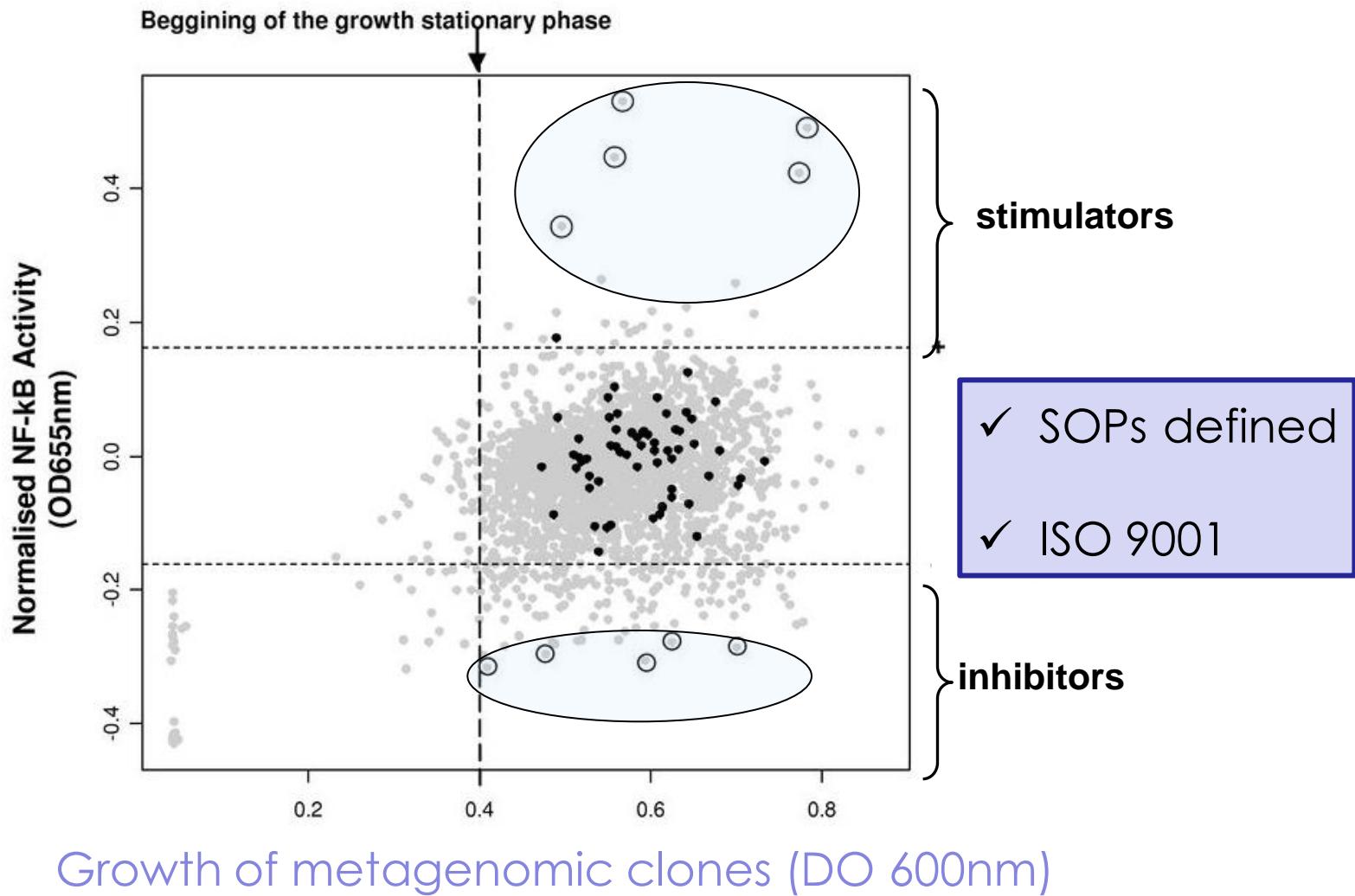
*Development of MetaFun*



Castor 1 & 2 – Human cells  
Pollux 1 & 2 – Bacteria culture  
Qpix – colony picker  
Cell sorter FACS ARIA III  
Automated Image Xpress confocal HCA



# HTS OF METAGENOMIC CLONES

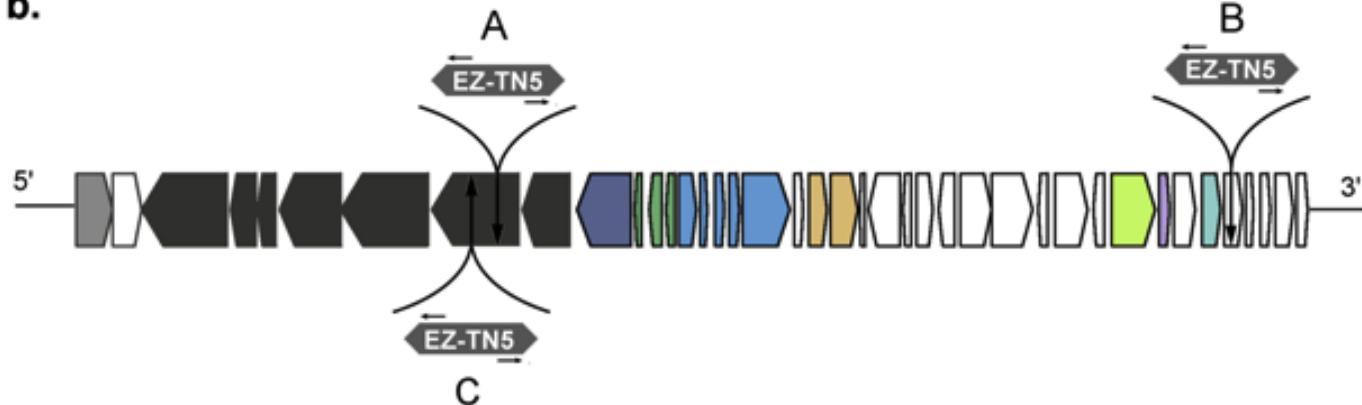


Lakhdari et al, PLoS One, 2010; de Wouters, PLoS one, 2014

## AN EXEMPLE : CLONE 52B7 (from Crohn 's Disease library)

**Stimulates NF-κB** in several cell lineages and **IL-8** secretion in IEC  
**Secreted** in the supernatant, size 50 KD, Trypsin sensitive  
From ***Bacteroides vulgatus***

b.

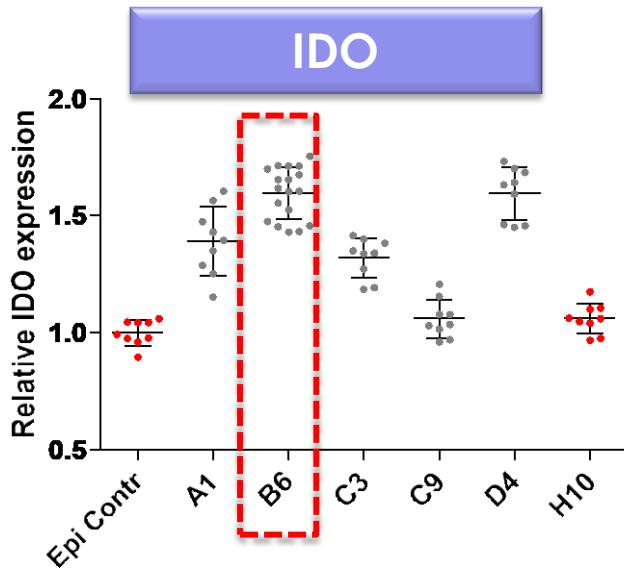
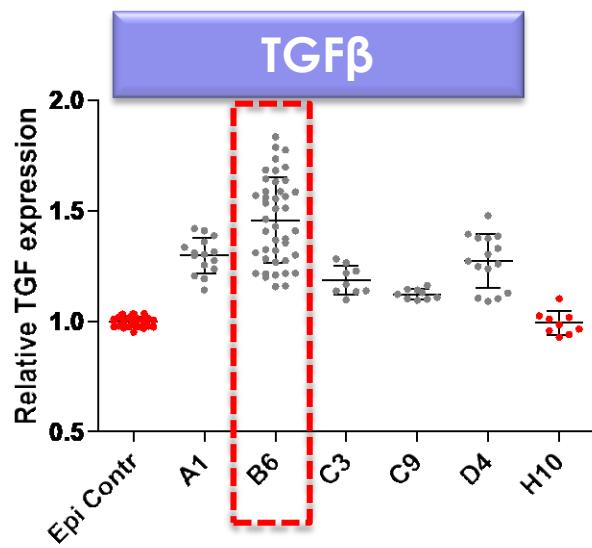


- ✓ The 2 targeted genes code for : a lipoprotein and a lipoprotein transporter
- ✓ Bacterial genes found more often in Crohn disease patients

Lakhdari et al, PLoS one, 2010, O'cuiv et al, Anaerobe, 2017

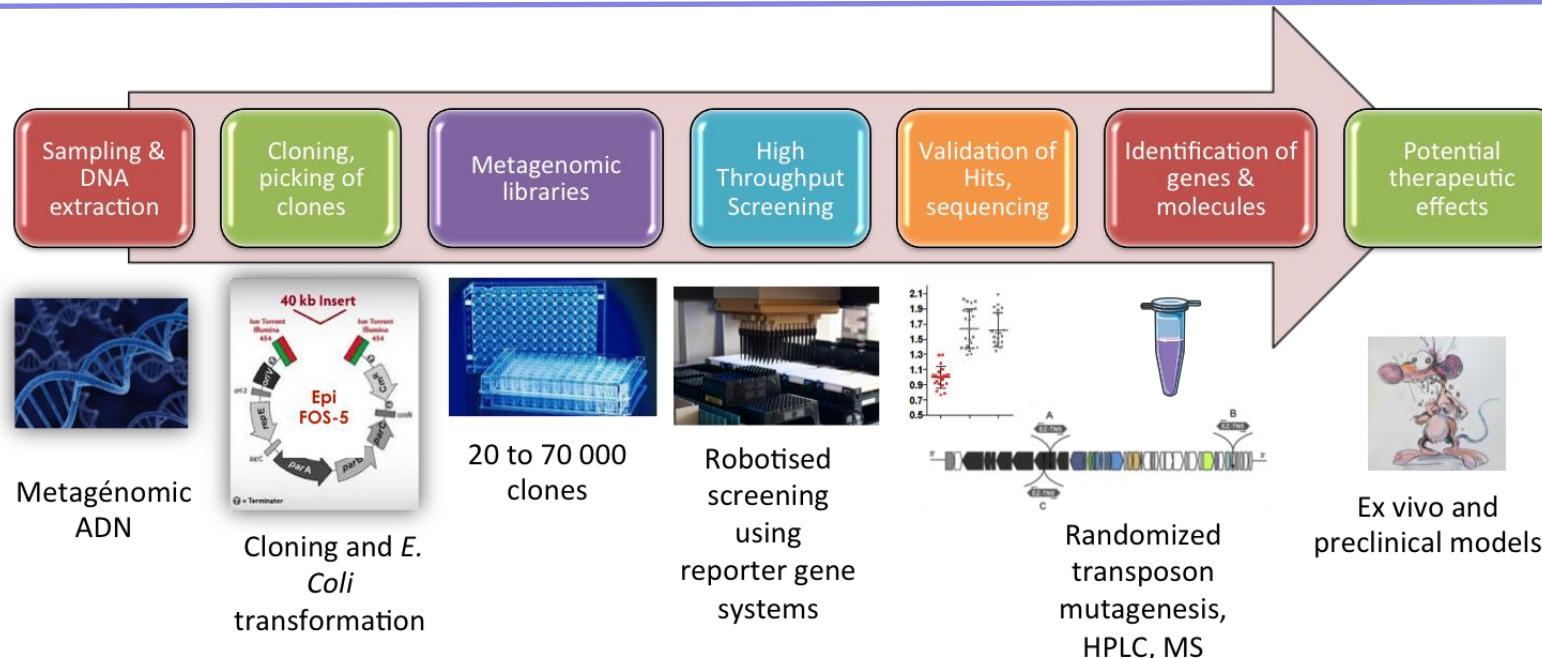
## ANOTHER EXAMPLE : CLONE B6 (from healthy's library)

- Stimulates NF-κB (MyD88 independent), IDO and TGFβ on colonocytes
- Secreted in the supernatant by a ABC transporter
- size < 1 kD
- Firmicutes: related to *Blautia* (*Clostridium XIVa*)



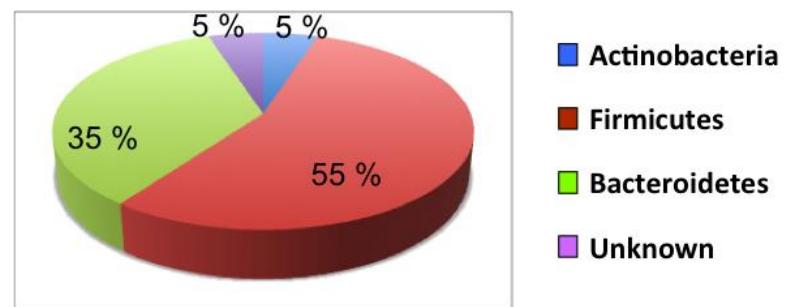
- ✓ One gene , Mate efflux family protein
- ✓ Mutagenesis on this gene: loss of function
- ✓ Metabolomic : NEW compound
- ✓ NEW PATHWAY/TARGET

# FUNCTIONAL METAGENOMIC : A FRUITFUL STRATEGY



>30 bioactive clones, identified with screens based on cell-assays on :

- Immunity
- Proliferation
- Metabolism



New molecules, new pathways discovered  
6 patents in 2015, licenced to

## TAKE-HOME MESSAGES

- ✓ Quantitative metagenomic allows functional understanding of microbiota host-interaction
- ✓ Animal models are useful tools but should be used with caution
- ✓ Functional metagenomics : A new strategy to study the molecular mechanisms of cross-talk between (uncultivated) gut microbiota and its host
- ✓ We are far from fully understanding microbiota-host cross-talk

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Catherine Juste,  
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Nicolas Pons  
Emmanuelle Le Chatelier  
V  ronique Lejard

# Merci

# Nous sommes pr  ts    travailler avec vous

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