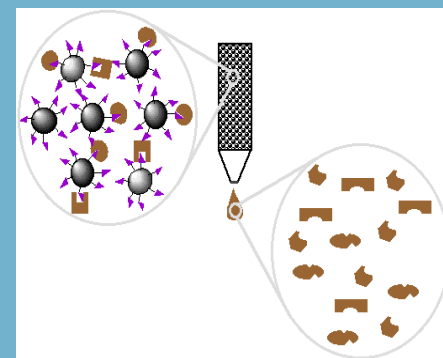
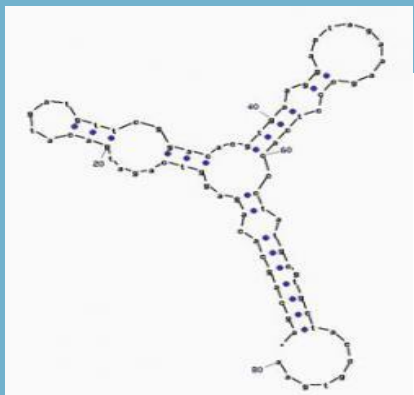


Aptamers: Powerful and Innovative Ligands in Affinity Chromatography

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Outline

1

Aptamers as affinity ligands: definition, selection and general properties

2

Selection of an Aptamer designed for affinity chromatography application

3

Examples of aptamo-purification

4

The path to industriability

5

Conclusions



Aptamers

(From Latin *aptus* : fitting, sticking to -)

Single-stranded nucleic acids that specifically recognize and bind tightly to their cognate targets due to their stable 3D structures

SCIENCE, VOL. 249 3 AUGUST 1990

Systematic Evolution of Ligands by Exponential Enrichment: RNA Ligands to Bacteriophage T4 DNA Polymerase

CRAIG TUERK AND LARRY GOLD

NATURE · VOL 346 · 30 AUGUST 1990

In vitro selection of RNA molecules that bind specific ligands

Andrew D. Ellington & Jack W. Szostak*

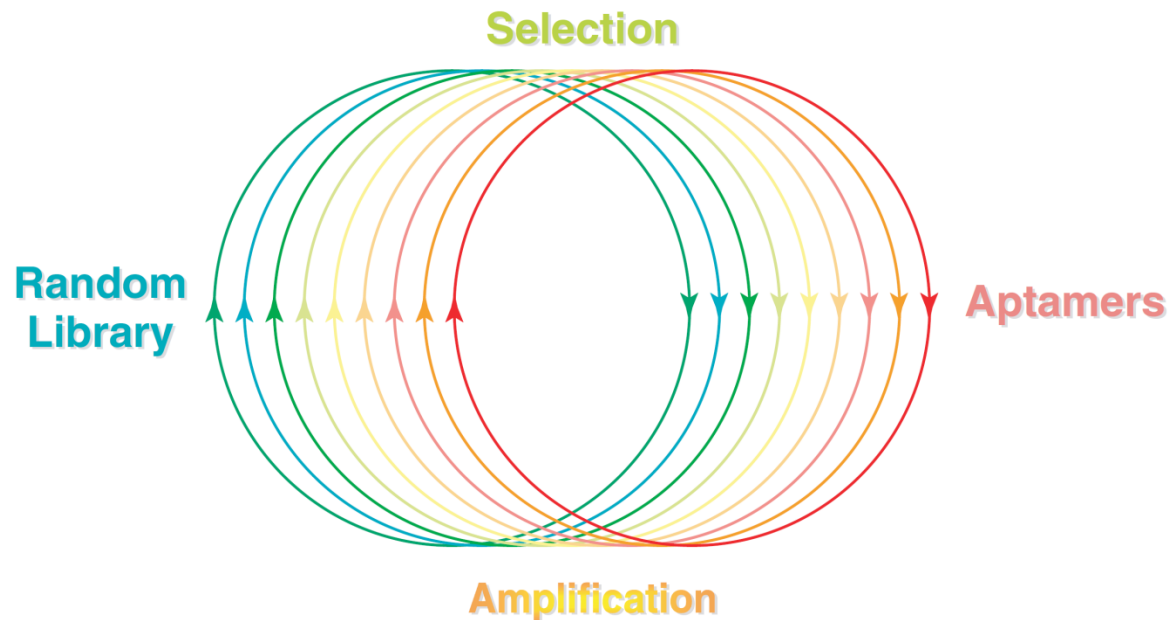
Department of Molecular Biology, Massachusetts General Hospital, Boston, Massachusetts 02114, USA



SELEX

Systematic Evolution of Ligands by Exponential enrichment:

An iterative *in vitro* process enabling the isolation, from a highly diverse combinatorial library of oligonucleotides, of the best binder(s) under a given set of conditions

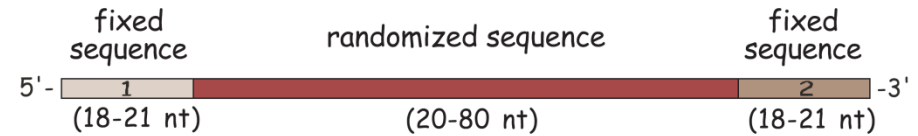


Toulmé et al., (2009) Aptamers: Ligands for All Reasons. In Aptamers in Bioanalysis

adebiotech IPP 2013



In vitro selection of target-specific aptamers using SELEX



$10^{13} - 10^{15}$
unique sequences

random
DNA oligonucleotide
library

in vitro
transcription

RNA library

Target

Binding

Partition

Elution

enriched pool
of selected
oligonucleotides

RNA
or
DNA

Conditioning

- in vitro transcription
- purification of the relevant ssDNA

SELEX
round

Amplification

- RT-PCR
- PCR

Last round of SELEX

Cloning

of the selected
aptamer pool

- sequencing
- sequence analysis
- binding studies
- post-SELEX modifications

individual
aptamers

RNA or DNA



Deconvolution of the Enriched Library

- Number of cycles needed for an efficient evolution of the library: 6 to 16
 - Stopped when the enrichment/affinity of the Aptamer pool is deemed sufficient enough
 - One SELEX leads to many specific candidates
- Sequencing performed on the evolved Aptamer pool allows
 - Identification of repeated sequence motifs & families
 - Characterization of their putative secondary structures and identification of the minimal core-sequences needed for binding



Aptamers : main attributes

- Binding to protein targets with K_d values usually in the low-nM to pM range
- Generation of aptamers against non immunogenic, or difficult to handle targets (Prions, toxins, small molecules)
- Quick *in vitro* process: Generation in ~ 4 - 8 weeks against virtually any target
- Owing to their small size: anti-protein aptamers can access/contact relatively small binding pockets on their targets
 - Powerful and highly specific inhibitors
- Can be selected under a wide range of conditions
 - Select species that bind under non physiological conditions (pH, ionic strength, solvent, temperature)
 - Binding under precise conditions in order to meet a specific need (process, type of sample)
- Their chemical nature (5'-P & 3'-OH) allows for precise derivatization (biotin, fluorescent reporter, spacer arm, etc.)



Applications

- Therapeutic agents
 - First FDA-approved Aptamer drug (Pegaptanib) for treating age-related macular degeneration (AMD)
 - Cancer: Adherence & Angiogenic factors, immune system modulators, Receptor Tyrosine Kinases (RTKs), and modulators of cell growth
- Molecular detection systems (Aptamer-based Biosensors: Aptasensors)
- Molecular Imaging
- Diagnostics & Biomarker discovery
 - SomaLogic Inc. & its SOMAmer-based Multiplexed Proteomic Technology
- Molecular targeting and drug delivery
 - Chimeric molecules containing an aptamer-conjugated with a small interfering RNA (siRNA) or short hairpin RNA (shRNA) that induce targeted RNA interference (RNAi)
-
- Affinity Ligands



Affinity ligand Ideal attributes ?

- Dedicated grafting function to allow oriented immobilization
- Capable to exclusively recognize the target protein
- Possibility to easily release the captured proteins by a chosen condition suitable for the product
- Reusable and stable under quite harsh conditions
- Neither leakage nor molecular hydrolysis
- Molecular mass as small as possible
- Non-toxic
- easy synthesis and affordable cost



Aptamer as ligand for affinity chromatography : Ideal ligand ?

■ High specificity and selectivity

- Generally considered to be superior to antibody-derived ligands

■ High physical and chemical stability: reusable and resistant to harsh sanitization

- High physical and chemical stability of DNA chemistry
- Modified nucleotides provide nuclease resistance

■ Behavior which may perfectly fit specific process requirements

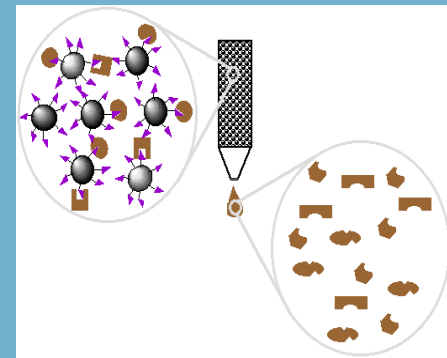
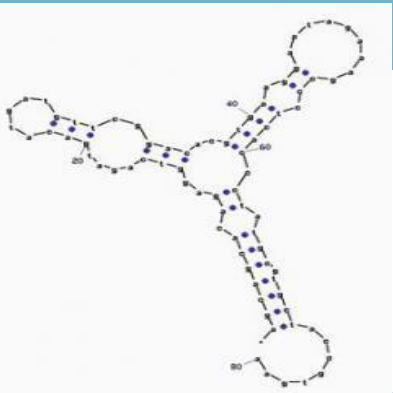
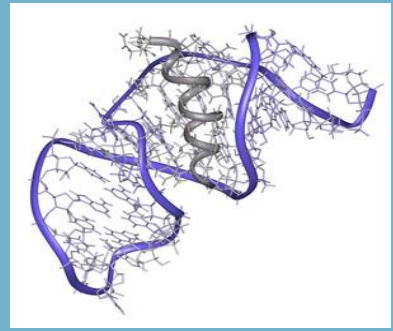
- Behavior chosen and modulated during the SELEX process
- Elution under optimal conditions for the product or considering the process constraints (modality of selection)
- Improvement selectivity through counter-selection
 - Particular interest for transgenic proteins: counter-selection against undesirable endogenous proteins



Aptamer as ligand for affinity chromatography : Ideal ligand ?

- Manufacturing cost: large scale production by chemical synthesis
 - High reproducibility and moderate cost: 2 to 5 K€ per gram of Aptamer with spacer and terminally-modified nucleotide (MW = 1/10 to 1/50 of Mab)
 - No potential biological contamination
- Availability of highly sensitive assays when considering aptamers as leachables
- Lack of immunogenicity (in case of leakage)
 - Lack of immunogenicity demonstrated during pre-clinical tests for therapeutic Aptamers when 1,000-fold higher doses were administered to monkeys (by Eyetec for anti-VEGF165).

Selection of an Aptamer designed for affinity chromatography applications





Selection of an Aptamer designed for affinity chromatography applications

Case of human Recombinant FVIIa

	SELEX strategy
Selection buffer	Tris 50mM / NaCl 50mM / CaCl ₂ 10mM / MgCl ₂ 4mM pH = 7.5
Target	Alternatively : R FVIIa / plasmatic derived FVIIa
Counter-selections	Support : Nitrocellulose filter Recombinant rabbit FVII (85% homology)
Wash / Elution	NaCl 2M / EDTA 10mM



Significant Proportion of interesting species in round 7 confirmed in round 10

→ Cloning and sequencing



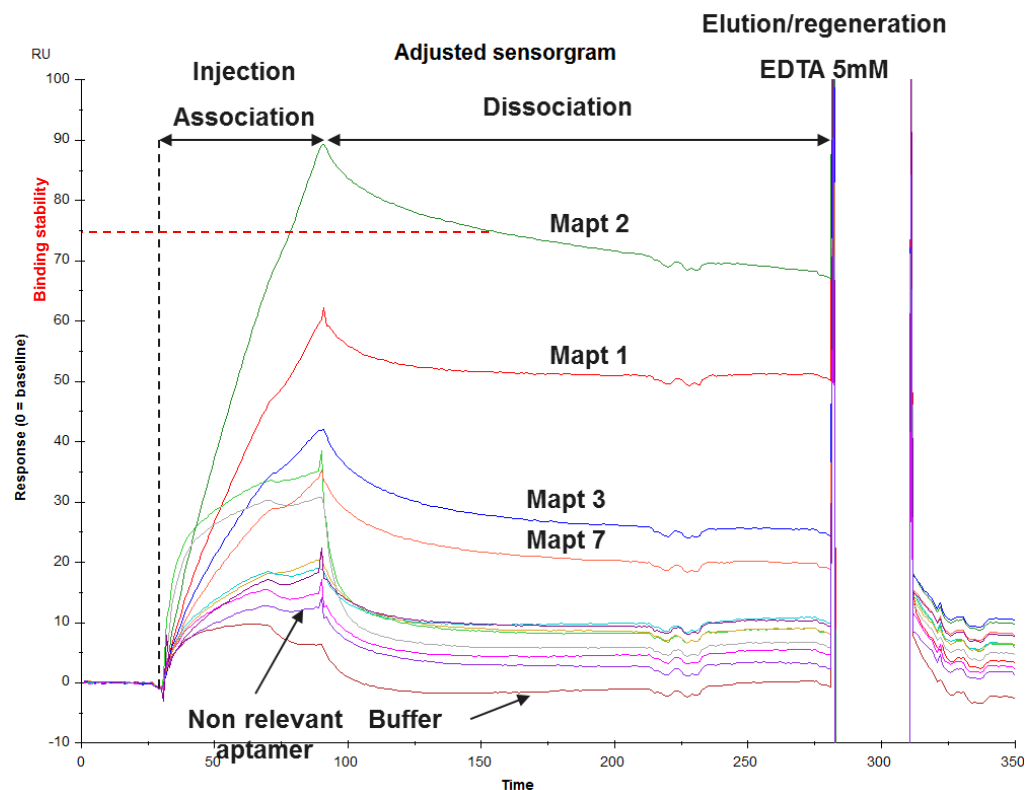
Selection of an Aptamer designed for affinity chromatography applications

Case of human Recombinant FVIIa

Bioinformatics and SPR analysis (Biacore) for confirmation of Monoclonal Aptamer (Mapt) sequences

	10	20	30	40	50
F10-S2T7TAAG	CCGCTGCGTA	TTATAGCTGA	ATGCC--C-	TAATGGGAAG TG
F12-S2T7
G11-S2T7C.T.	A C
E03-S2T10C.	A
E09-S2T10C.	C
A03-S2T7CC.C.	A
E01-S2T7C.A.	A
E08-S2T10C.T.	T..TT
G09-S2T10C.	T..TT
H07-S2T10CC.	G.....	C.....	T..T	C..T
B08-S2T10C.T	T..CT
G5-S2T7C.	T..A	CT..T
E10-S2T10	GTGCAGCC.	A.TN.	AGTG.	T..AAG.	A.G.G.---
A9-S2T10	...ATGC.	-AGC..C.	G.G..AG	AGA.A	GG.C-T.
B01-S2T10CC.	A.AGC..C.	G.T.CAG	AGAT.A	GG.C-T.
Clustal Co	*		***	***	*

Bioinformatics alignment for family identification



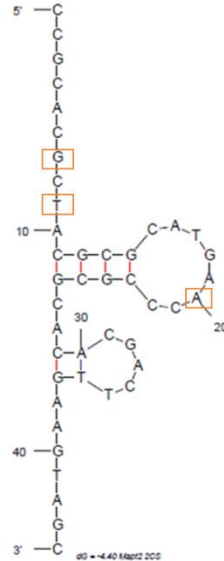
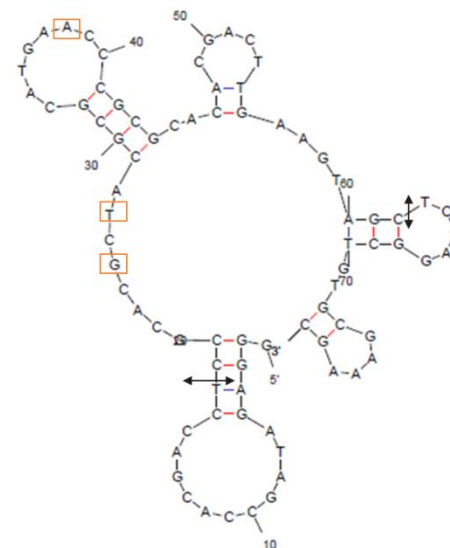
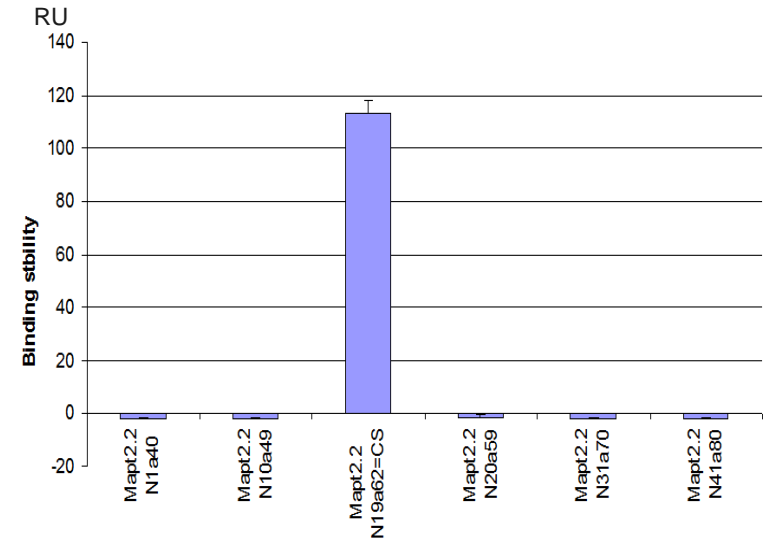
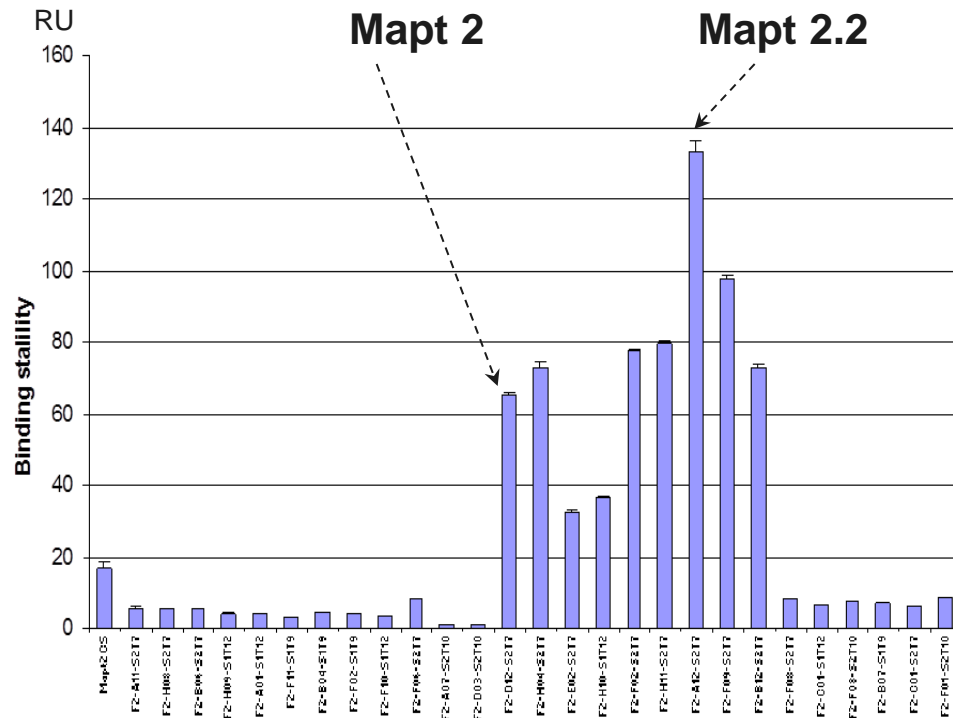
Affinity confirmation against immobilized target : R FVIIa



Selection of an Aptamer designed for affinity chromatography applications

Case of human Recombinant FVIIa

SPR analysis for best binder determination in a aptamer family followed by core sequence identification



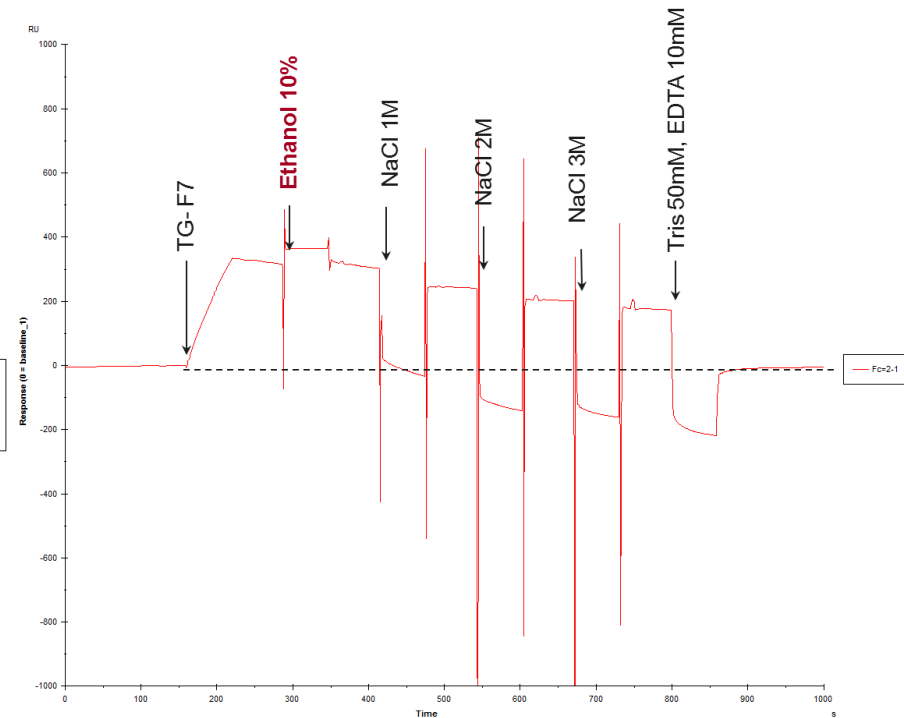
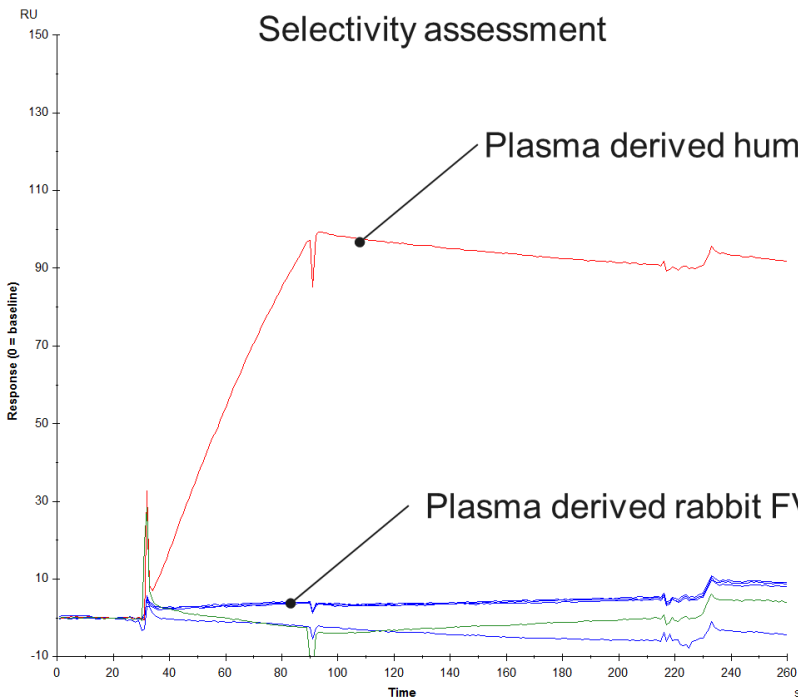
Orange box: Mutation with high impact on affinity



Selection of an Aptamer designed for affinity chromatography application

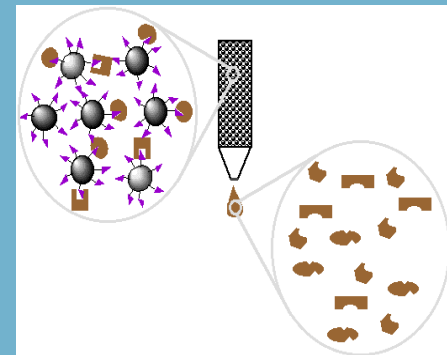
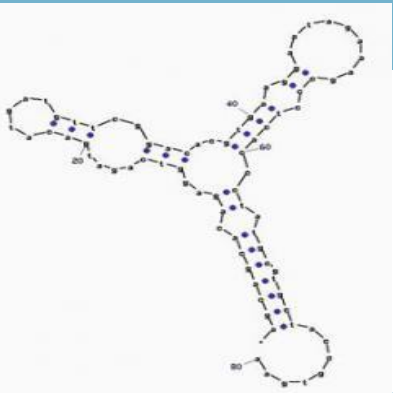
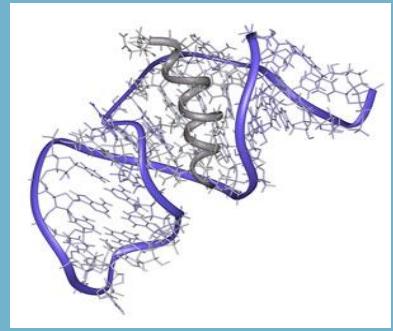
Case of human Recombinant FVIIa

Selectivity assessment and analyses of their binding properties



Resistance under different wash conditions
evaluation with immobilized aptamer

Examples of Aptamo-purification



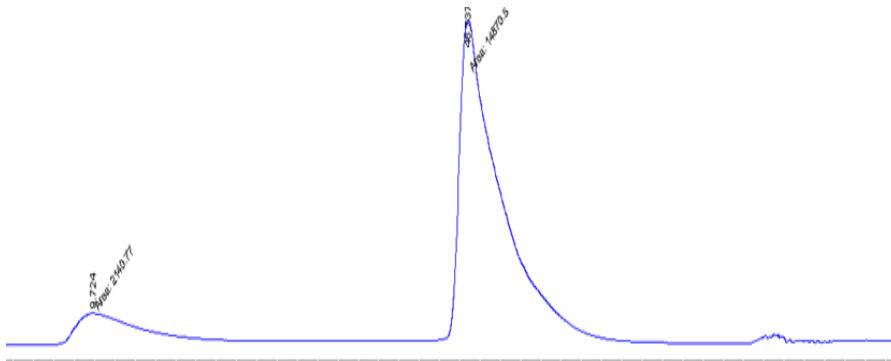


Aptamo-purification as a polishing step

Example of Recombinant FVIIa

From highly purified material

Purified recombinant FVIIa from slightly stressed raw material containing 10% of product-related impurities (other impurities less than 10ppm)



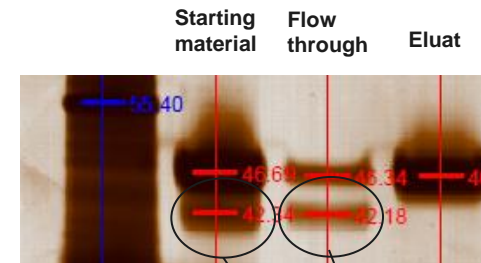
Material: Purified hRFVIIa

Equilibration buffer:

50 mM Tris-HCl (pH 7.4)
150 mM NaCl
10 mM CaCl₂

Washing Buffer: none

Elution: 50 mM EDTA pH 8



SDS PAGE gel - silver staining

Apparent molecular weights indicated

⇒ Increased specific activity (ratio of chromogenic activity to antigen titer) from 0.7 to 1.0

Identified by MS as des-Gla FVII

⇒ Capacity to eliminate very closely related impurities



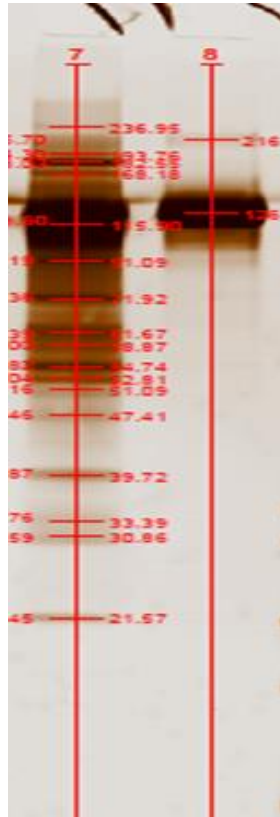
Highly purified product by aptamo-purification

Example of Factor H protein

From **pre-purified** material

Plasmatic FH obtained after 4 chromatographic steps, including a pseudo affinity chromatography

Starting material Eluate



From **crude** material

Recombinant FH in supernatant

Starting material Eluate



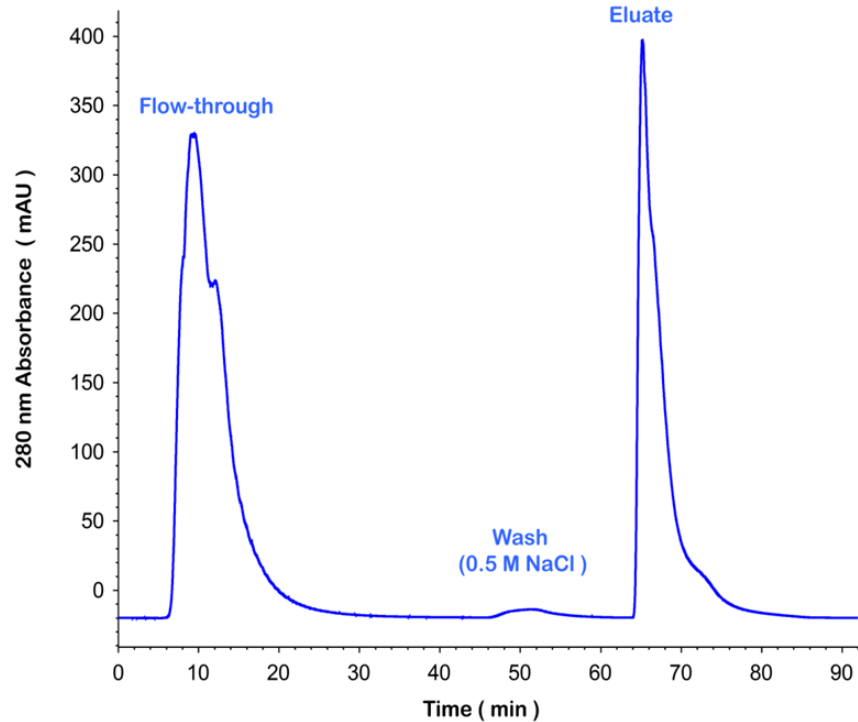
➔ Highly purified FH product obtained in on single step (with 1M NaCl wash) from pre-purified or crude material



Highly purified product by aptamo-purification

Example of coagulation Factor IX

Nonapta5.1 grafted – 1 mL resin



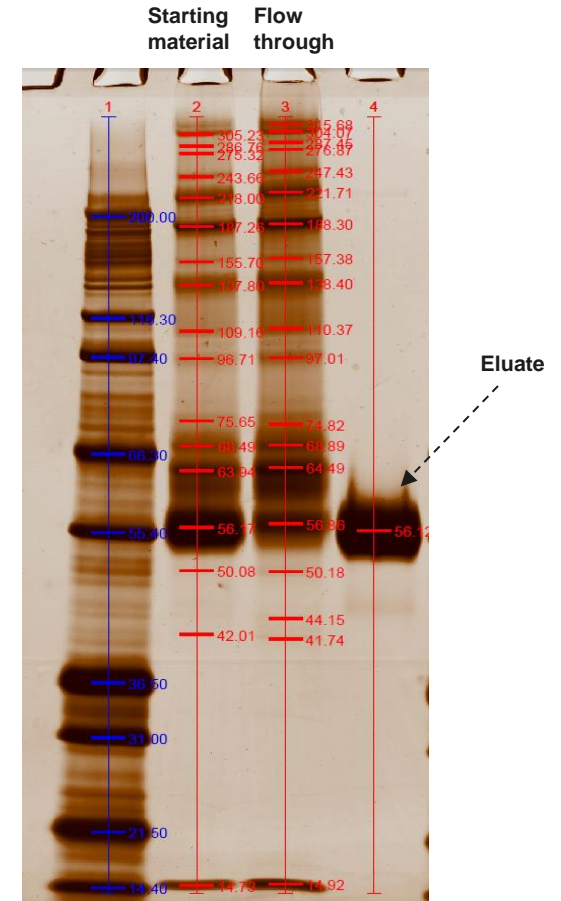
Material: Intermediate product containing 94 UI/mL of FIX

Equilibration buffer:

50 mM Tris-HCl (pH 7.4)
150 mM NaCl
2 mM CaCl_2
1 mM MgCl_2

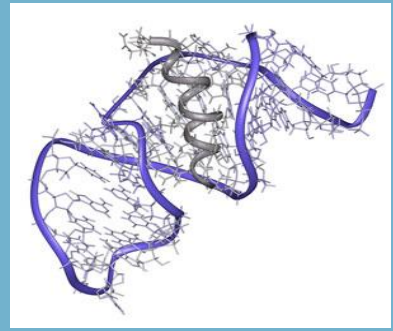
Washing Buffer: Equilibration buffer with 0.5 M NaCl

Elution: 200 mM EDTA (pH 8)

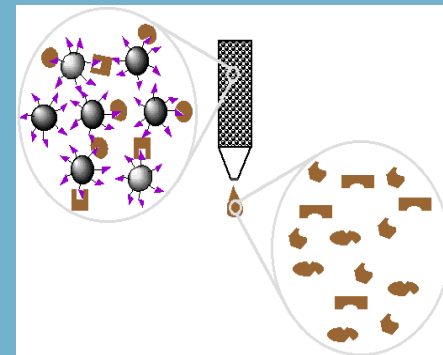
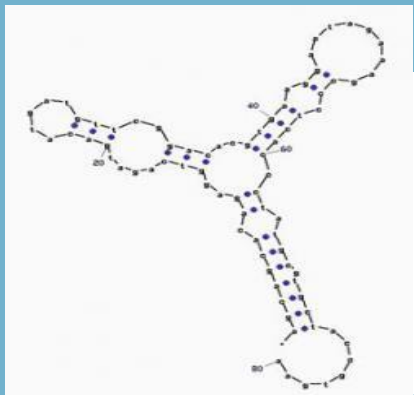


SDS PAGE gel - silver staining

Apparent
molecular weight indicated



The path to Industriability: Grafting and resistance demonstration



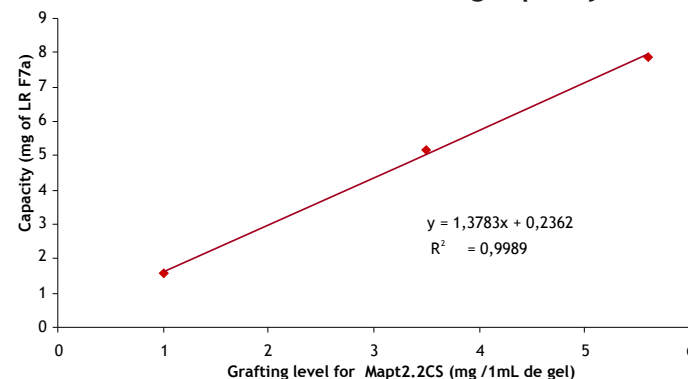


On the path to Industriability: High grafting yield

A specific concern for this technology has been addressed through the use of a proprietary immobilization approach (efficient chemical grafting of an NH₂-derivatized oligonucleotide on a classical support)

Quantity targeted in µg (for 1mL gel)	100	3500	6000
Quantity grafted in µg	≈ 100	3498	5556
Grafting yield (%)	100.0	100.0	92.6

Immobilization level/binding capacity relationship



Immobilization (mg)	1.0	3.5	5.7
Dynamic Capacity* (mg)	1.6	5.2	8.0

*Capacity for a 50 KDa protein at 60 cm/H

- ⇒ New grafting approach which provides a very high yield of immobilization
- ⇒ High efficiency (functionality/grafting)

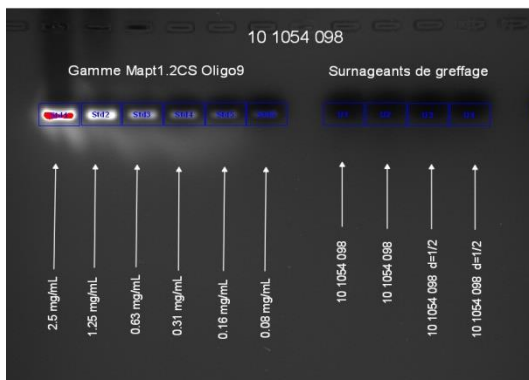


On the path to Industriability: Specific and sensitive detection assays

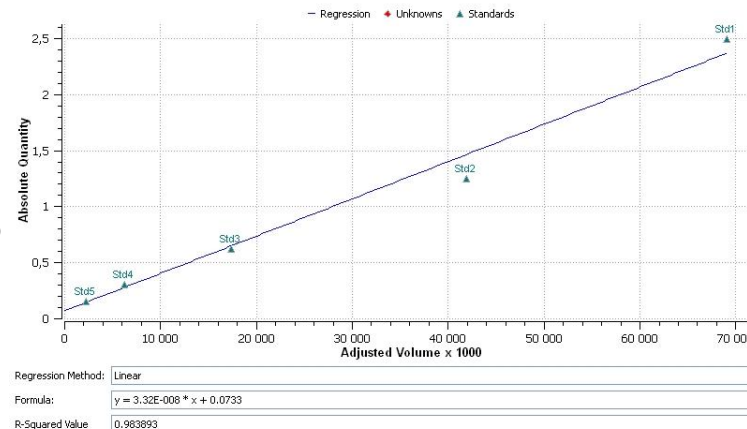
- For most of affinity ligands, the development of a specific assay especially in the presence of the target remains a challenge
 - Sensitivity required less than pg/mL
 - Overcomes the probable complex formation with the target and/or the probable matrix effect by the concentrated target itself in the final product
- For aptamers different methods developed in the genomic and molecular biology fields can be adapted for this purpose
 - Highly sensitive methods available from molecular biology for DNA assay including a specific DNA extraction and/or target denaturation

On the path to Industriability: Specific and sensitive detection assay

- For most of affinity ligands, the development of a specific assay especially in the presence of the target remains a challenge
 - Sensitivity required less than pg/mL
 - Overcomes the probable complex formation with the target and/or the probable matrix effect by the concentrated target itself in the final product
- For aptamers different methods developed in the genomic and molecular biology fields can be adapted for this purpose
 - More simple approach to handle higher quantities (i.e. for grafting yield calculation): agarose gel analysis



Grafting yield analysis in agarose gel stained by SYBR® safe



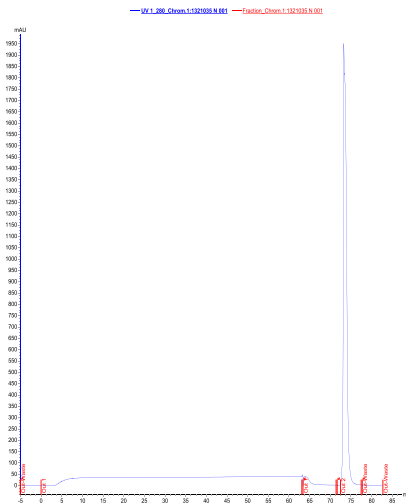
On the path to Industriability : Resistance of the aptamo-affinity resin

As a model : chemically immobilized anti-FVII aptamers with inverted 3'T

⇒ 2 anti-FVII aptamers with 2 different spacers (hyrophobic C6 and hydrophilic C11)

Parameters monitored:

- Capacity at 80% = 6.5-7.5 g/L (initial values)
- Specific activity of the eluted product : 0.8-1.0 (initial values)



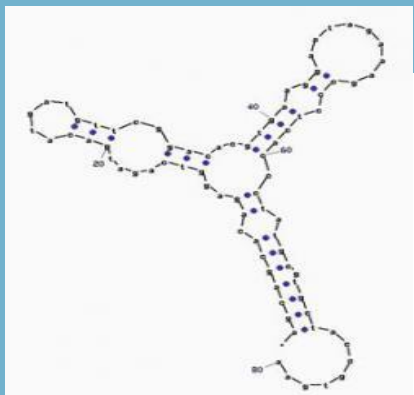
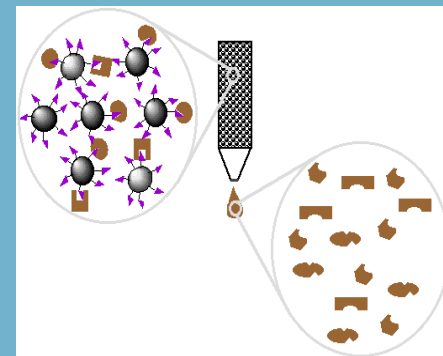
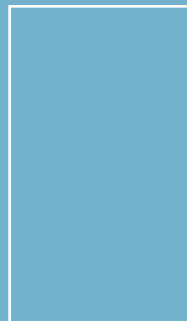
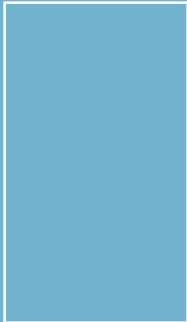
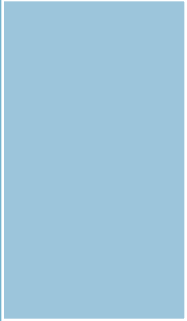
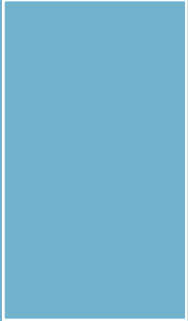
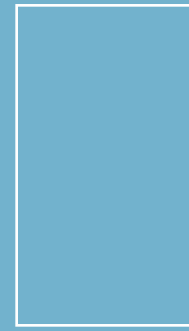
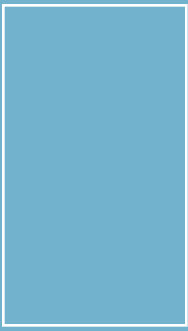
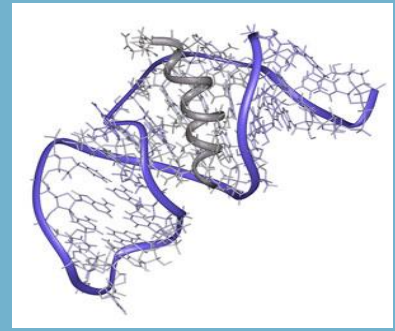
Typical chromatogram
obtained on pre-purified
product

Résistance tested	Mapt 2 C11		Mapt 2,2 C6	
	Capacity 80% g/L	Specific activity (FVIIam/FVIIAg)	Capacity 80% g/L	Specific activity (FVIIam/FVIIAg)
100 Hours Sodium hydroxide 1 M	7.1	0.9	6.9	0.9
100 Hours in cryosupernatant	7.3	0.9	ND	ND
100 Hours in clarified milk	ND*	ND	7.1	0.9
+ 30 add. cycles buffer/ sodium hydroxyde 1M	ND	ND	6.9	1.0

*not determined

⇒ Very high resistance of aptamer resin

Conclusions





Aptamer for affinity chromatography

A major interest for LFB : Aptapure®

- Most promising technology for improving yield and COGs of biotherapeutics including Plasma fractionation
 - 9 patents submitted
 - One major patent: original process for Aptamer immobilization
- The last 3 years, investment made by LFB aimed at demonstrating :
 - High selectivity (vs contaminants, related-protein contaminants, homologous proteins)
 - Capacity to discriminate proteins with correct PTMs (i.e. complete γ -carboxylation for FIX)
 - High resistance: ultimate sanitization with 1M NaOH, compatibility with biological media (serum, milk)
 - High yield of immobilization using a specific chemistry on a classical support
 - High efficiency of immobilized aptamers
 - Very sensitive assays of residual aptamers in case of leakage
- First positive feedback from EMA (Innovation Task Force)



Special Thanks

- Sami Chtourou, Patrick Santambien, Laurent Siret and Nicolas Bihoreau
- My team: Cynthia Forier, Agnès Cibiel, Clément Costa, Alexander Seifert and Mohamed Ouhammouch
- Process team : Michel Nogré, Michel Tellier and Damien Bataille
- Egisto Boscheti (Jam Conseil) and Frédéric Ducongé (INSERM-CEA)

Thank you for your attention !